Bacteriological and Biochemical Studies On Pekin Duckling Infected With Pasteurella Multocida with Trial For Treatment

Allam HH¹, Aly Salah ABS², Zaki R H³ and Nagwa A Bahar⁴ (Poultry¹ and Biochemistry (2,3 and 4) Department)

Animal Health Research Institute (Zagazig (1 and 3), Kafer El Sheik and Ismailia Branches)

ABSTRACT

Samples from cloacal swabs, liver, heart, lungs, trachea, spleen and nasal exudate were collected from 150 pekin duckling ageing 1-30 days (75 apparentaly healthy 35 diseased and 40 freshly dead) for bacteriological examination. Out of 150 examined sample 43 (28.67 %) were positive for Pasteurella Multocida namely 5 from apparentally healthy, 13 from diseased and 25 from dead ducklings. Antibiogram study of isolates revealed that florfenicol was the drug highly effective against Pasteurella Multocida

A total of 160, one day old pekin duckling (80 healthy & 80 experimentally infected with Pasteurella Multocida) were divided into 4 equal groups (40 birds each), the 1st group consisted of healthy ducklings (control), the 2nd group inculded healthy ducklings were treated with florfenicol (30mg/kg bwt.) in drinking water for 5 days, the 3rd group inculded infected non treated and the 4th group inculded infected and treated with florfenicol (same dose, period and route of administration). In the four groups the hemato-biochemical changes were studied.

The results revealed that healthy pekin ducklings treated with florfenicol displayed significant rise in body weight, leukocytic count, lymphocytosis, significant decrease in heterophils and insignificant increase in monocytes, eosinophils, basophils, total proteins, albumin, globulins, A/G ratio, AST, ALT, ALP, uric acid and creatinine.

Pasteurellosis in duckling induced anorexia depression, ruffled feathers, coughing, diarrhea, rise respiratory rate and 30% mortality rate. Monocytes, total, β, γ globulin, AST, ALP, uric acid, creatinine levels and significant decrease in weight gain, heterophils, albumin, a globulin, A/G ratio, insignificant decrease in leukocyte, lymphocyts, esinophils, total proteins and insignificant rise ALT

Florofenicol residues in examined liver and kidneys in both treated healthy and diseased ducklings were high at 1st day post treatment, very low at 6th day and completely negative from examined samples at 9th day post treatment. The highest levels of florofenicol residues were recorded in kidneys then liver.

Duckling suffering from pasteurellosis and treated with florfenicol showed no clinical signs, mortality rate 5%, reduced reisolation Pasteurella Multocida and Improved hemato-biochemical parameters.

It could be concluded that florfenicol is effective in treatment of Pasteurella Multocida in duckling

INTRODUCTION

Ducks are relatively resistant to some diseases (1). In duck farms in Egypt, poor management, inadequat diagnosis, control and

prevention of various infectious diseases play a vital role in high morbidity and mortality (2). Pasteurellosis is a contagious disease in ducks caused by Pasteurella Multocida (3) which is

associated with poor sanitation (4). Acute form occurs as septicemia of sudden onset with high mortality (5). While chronic form results in localization of infection in wattles, respiratory passages and joints (6). Symptoms of pasterulosis include aneroxia, mucous discharge from mouth, diarrhea and laboured breathing (7).

Florfenicol is a novel broad spectrum bacteriostatic antibiotic belonging to family including also thiamphenicol and chloramphenicol (8). It has fluorine atom instead of hydroxyl group located at C-3 in the structure of chloramphenicol and thiamphenicol (9). Florfenicol inhibits protein synthesis (10), it has greater activity than chloramphenicol and thiamphenicol against pasteurella (11).

The aim of this study was to evaluate effects of treatment in experimentally infected ducklings. Biochemical parameters.

MATERIAL AND METHODS

Isolation and identification

A total of 150 samples (35 diseased, 40 freshly dead, 75 healthy pekin duckling) 1-30 days old were obtained from different private farms at Sharkia Province. All samples were aseptically collected from cloacal swabs, liver, heart, lungs, trachea, spleen and nasal exudate and inoculated into nutrient broth aerobically at 370C over night, subculturing on blood agar and MacConkey agar plates was performed for 24h at 370C, suspected colonies were identified (12). Pathogenicity and virulence of isolated P milt to mice were determined (13).

Antibiotic sensitivity test

Sensitivity of Pasteurella Multocida to different chemotherapeutic agents was tested by disc diffusion method (14).

Drugs

Florfenicol (Aviflor, 100 mg/ml) water soluble formulation for oral use was supplied by Avico (Jordan).

Experimental duckling

A total of 160, one day old pekin duckling were reared under hygienic conditions, Fed on balanced commercial starter ration free from antibacterial agent and water ad-libitum,

Pasterulla multocida infection

On day 30 of age 80 duckling were I/M inoculated with 0.2 ml/bird of 48 hr broth culture of pasterulla multocida. containing (3x108CFU) viable organism (15).

Experimental design

Duckling were divided into 4 equal groups (40 ducks each), 1st group inculded non-infected non-treated duckling (control group), 2nd group inculded non-infected duckling treated with florfenicol (30 mg/kg bwt) in drinking water for 5 successive day. The 3rd group inculded infected non treated duckling and the 4th group inculded infected duckling treated with florfenicol (same dose, period and rout of administration). Treatment started at age of 32 day.

Body weight

From each group 5 ducklings were weighted individually at the start of the experiment and at the 1st day post treatment and consumed diets were record, calculation of weight gain and feed conversion rate.

Sampling

At 1st, 7th and 15th days post treatment two blood samples from all ducklings were taken, the 1st sample was taken in tube containing EDTA to estimate leukogram (16). The 2nd one was taken to obtain serum to estimate total protein (17). Protein fraction was performed using cellulose acetate electrophoresis (18). (AST-ALT) (19) ALP (20) uric acid (21) creatinine (22).

Re-isolation of Pasteurella Multocida

Samples were taken aseptically from cloacal, nasal exudates and internal organs from all groups post treatment then inoculated into nutrient broth at 370C over night followed by subculturing on nutrient agar for 24h at 370C, suspected colonies were identified (12).

Drug residues

Five ducklings from healthy treated and infected treated ducklings were slaughtered at 1st, 3rd, 6th& 9th days post treatment. Levels of florofenicol residues in liver and kidney were

determined by microbiological assay technique (23) using Baceillus subtilis as test organism Statistal Analysis

Obtained data were statistically analyzed using T test (24).

RESULTS AND DISCUSSION

Table 1. Incidence of the isolated Pasterulla multocida from ducks

Source of sample	Total No. of	+ve	samples	-ve samples	
	sample	No.	%	No.	%
Apparently Healthy duckling	75	5	6.67	70	93.33
Diseased duckling	35	13	37.14	22	62.86
Freshly dead duckling	40	25	62.50	15	37.50
Total	150	43	28.67	107	71.33

Table 2. Incidence of the isolated Pasterulla multocida from internal organs in duckling

Source of	Не	Healthy			Disased			dead		
sample	Total	+ve	sample	Total	+ve s	sample	Total	+ve	sample	
	sample	No	%	sample	No	%	sample	No	%	
Tracheal swab	35	3	8.57	5	2	40	8	4	50	
Cloacal swabs	40	2	5	3	1	33.33	6	3	50	
Liver	-	-	-	9	2	22.22	7	4	57.14	
Heart	-	-	-	6	2	33.33	3	3	100	
Lung	-	_	_	3	2	66.67	9	6	66.67	
spleen	-	-	_	5	1	20	7	5	71.43	
nasal exudates	-	-	-	4	1	25	-	_	-	
Total	75	5	6.67	35	13	37.14	40	25	62.5	

Table 3. Effect of Pasterulla multocida on mortality rate and reisolated Pasterulla multocida of duckling

	Parameter: Total		Mortality rate		Reisolation	
Group	No	Signs	No	%	No	%
Healthy non treated (Control)	40	00	00	00	-	0%
Healthy Florofenicol Treated	40	00	00	00		0%
Diseased non treated	40	38	12	30	40/40	100%
Diseased treated	40	2	3	5	3/40	7.5%

Bacteriological examination revealed that 43 samples (28.67%) out of 150 collected samples to be positive for Pasterulla multocida (5 from healthy, 13 from diseased and 25 from freshly dead duckling). Pasterulla multocida was isolated from diseased duckling in a percentage of 25% (25).Experimental

pasteurellosis in duckling induce clinical signs mainly, depression, anorexia, ruffled feathers, increased respiratory rate, sneezing, coughing, diarrhea, mortality rate 30% and decrease body weight. Typical signs were previously recorded in duckling (26).

Table 4. In vitro susceptibility Pasterulla multocida to some commonly used antibiotics

Antimicrobial agent	Disk polancy	Inhibition zone diameters(mm)	Sensitive
Florphencol	30ug	25	+++
Ceftiofur sodium	30ug	25	+++
Gentamycin	10ug	23	+++
Spectinomycin	10ug	22	++
Colistine	30ug	16	R
+++ = high sensitive	++ = sensiti	ve $R = Resistance$	

Antibiogram revealed that Pasterulla multocida was sensitive to florfenicol and ceftiofur sodium followed by gentamycin, spectinomycin and resistant to colistin. Same

reports showed that Pasterulla multocida were very sensitive to florfenicol (25). Other report that Pasterulla multocida were resistant to colistin (27).

Table 5. Effect of Pasterulla multocida and florofenicol on body weight gain (gm) and feed conversion rate in duckling (n=5)

Parameter	Control group	Healthy treated	Diseased Non treated	Diseased treated
Weight at 30 th day of age	410.32±1.49	418.58±1.37	405.08±1.40	415.41±1.86
Body weight at 1 st day PT	480.49±5.38	502.04±9.80*	464.22±3.50*	489.55±15.94
Weight gain	69.45± 1.98	83.46±3.97*	59.14± 2.13*	74.14± 1.85
Feed consumption	238.59	243.39	217.43	228.53
Feed consumption rate	3.45	2.92	3.68	3.08

*Significant at P ≤ 0.05

PT= post treatment

The obtained results revealed that. florfenicol treatment in healthy ducklings displayed a significant increase in body weight gain in comparasion to healthy duckling, meanwhile; pasteurellosis induced significant decrease in body weight gain and feed

conversion rate in comparasion with healthy ducklings. The antimicrobials drugs induce rise in growth rate through inhibiting pathogenic organisms (28). Pasteurellosis in duckling induced significant decrease in body weight gain and increase in feed conversion rate (29).

Table 6. Effect of *Pasterulla multocida* and florofenicol on leukogram in duckling (n=5)

	Control	Healthy	Diseased					
Parameter	group	florofenicol	Non	Day	Day post treatment			
		treated	Treated	1st	7th	15th		
$TLC(X10^3)$	15.87±	17.31±	14.90±	15.06±	15.63±	15.75±		
TEC(MIO)	0.50	0.27*	0.15*	0.92	0.99	0.65		
Heterophils	4.12±	2.15±	2.59±	$3.07 \pm$	$3.84 \pm$	4.08±		
	0.85	0.19*	0.37*	0.40*	0.63	0.49		
Expmphocytes	8.09±	$10.90 \pm$	$7.40 \pm$	7.51±	7.77±	$8.02 \pm$		
Extra C rymphocytes	0.64	0.98*	0.89	0.78	0.56	0.60		
Eosinophils Basophils Basophils	1.34±	1.57±	$1.07 \pm$	1.10±	1.25±	$1.30 \pm$		
Eosmophins	0.35	0.41	0.55	0.70	0.69	0.50		
Basophils	$1.17 \pm$	1.33±	1.16±	1.18±	1.18±	1.15±		
— O Dasobinis	0.22	0.35	0.32	0.62	0.71	0.62		
Monocyte	1.15±	1.36±	$2.68 \pm$	2.20±	1.59±	1.20±		
wionocyte	0.30	0.34	0.57*	0.61*	0.54	0.93		

^{*}Significant at $P \le 0.05$

Florfenicol induced significant leukocytosis, lymphocytosis, significant decrease in heterophils, insignificant rise in monocytes, eosinophils and basophils. Pasteurellosis in ducklings induced insignificant decrease in leukocytic count, lymphocyts, esinophil, significant decrease in heterophils and monocytosis in duckling. Same

results were previously reported in healthy rats treated with florfenicol for 6 days (30). Our result agreed with those obtained in broiler chicken infected with Pasterulla multocida (31). Pasteurellosis in ducklings induces insignificant reduction in lymphocyts beside significant decrease in heterophils and monocytosis (26).

Table 7. Effect of Pasterulla multocida and florfenicol on protein profile in duckling (n=5)

		Control	Healthy	Diseased					
Parameter		group	florofenicol	Non	Day post treatment				
,			treated	Treated	1st	7th	15th		
T. protein	n (g/dl)	6.09±	6.87±	5.95±	6.03±	6.08±	6.13±		
		0.10	0.14	0.18	0.22	0.15	0.19		
Albumir	ı (g/dl)	$3.83 \pm$	4.35±	3.13±	3.41±	$3.60 \pm$	$3.79 \pm$		
		0.42	0.60	0.28*	0.16*	0.13	0.15		
	α	$0.70 \pm$	$0.77 \pm$	$0.49 \pm$	$0.54 \pm$	$0.68 \pm$	$0.73 \pm$		
Globulin		0.19	0.13	0.10*	0.08*	0.17	0.21		
(g/dl)	β	$0.83 \pm$	$0.95 \pm$	1.25±	1.20±	$1.00 \pm$	$0.86 \pm$		
		0.09	0.16	0.12*	0.07*	0.12	0.17		
	γ	$0.73 \pm$	$0.80 \pm$	$1.08 \pm$	$0.88 \pm$	$0.80 \pm$	$0.75 \pm$		
		0.10	0.18	0.11*	0.09*	0.14	0.15		
	Total	2.26±	2.52±	$2.82 \pm$	$2.62 \pm$	2.48±	$2.34 \pm$		
		0.37	0.91	0.14*	0.19*	0.17	0.18		
A/G R	atio	1.69±	1.72±	1.12±	1.30±	1.45±	1.62±		
		0.15	0.42	0.20*	0.19*	0.25	0.33		

^{*}Significant at P ≤ 0.05

In the current work, florfenicol displayed insignificant increase in total proteins, albumin globulins. Pasteurellosis in ducklings showed insignificant hypoproteinemia, significant rise in total, β , γ globulin, significant hypoalbuminemia, α globulin and A/G ratio. Simialar results were recorded in healthy broiler chicken treated with florfenicol (32-33). Changes in protein profile in ducklings suffering from pasteurellosis agreed with (34) in duckling and

(35) in chicken. Reduction in albumin in duckling infected with Pasterulla multocida may be due to effect of bacteria and its toxin in liver (sole of albumin synthesis) (36). Rise in β and γ globulin indicate activation of immune system due to infection (37). Reduction in globulin in infected duckling indicating immune defense mechanism against infection and enhanced immunoglobulin synthesis (38).

Table 8. Effect of Pasterulla multocida and florfenicol on liver enzymes and kidney function in duckling (n=5)

D	Control	Healthy		Diseased			
Parameter	group	group florofenicol		Day post treatment			
	172	treated	Treated	1 st	7 th	15 th	
AST	30.28±	32.04±	36.04±	34.78±	33.20±	31.08±	
$((\mu/L)$	1.50	1.78	1.41*	1.17*	1.44	1.48	
ALT	24.05±	25.34±	29.13±	27.85±	26.30±	24.97±	
$((\mu/L)$	1.42	1.89	1.89	1.47	1.60	1.93	
ALP	$17.09 \pm$	19.21±	21.87±	21.03±	19.14±	18.21±	
(I.U/ml)	1.12	1.96	1.04*	1.10*	1.79	1.43	
Urea	4.92±	5.58±	7.88±	7.61±	6.05±	5.12±	
(mg/dL)	1.22	1.60	1.10*	1.03*	1.02*	1.40	
Creatinine	1.14 ±	1.25 ±	1.69 ±	1.54 ±	1.34 ±	1.40 1.20 ±	
(mg/dL)	0.11	0.21	0.17*	0.09*	0.13	0.15	

Healthy duckling treated with florfenicol revealed insignificant increase in AST, ALT ALP, uric acid and creatinine. Pasteurellosis in duckling induce significant rise in AST, ALP, uric acid and creatinine levels beside insignificant rise ALT. Same results were obtained in healthy chickens treated with florfenicol (39). Florfenicol had no adverse

effects on liver and kidney functions (40). Same changes in uric acid and creatinine in duckling suffering from pasteurellosis were previously recorded (41). Rise in enzyme activities may be due to liver damage by infectious agent and its toxins (42). Elevation in uric acid and creatinine could be due to effect Pasterulla multocida or its toxin on kidney (43).

Table 9. Florfenicol residues (µg/gm) in duckling liver and kidney

	Clearance period										
	Healthy d	uckling witl	n florfenicol	(days)	Diseased (duckling wit	th florfenico	(days)			
	Ist	3rd	6th	9th	1st	3rd	6th	9th			
Kidney	6.72±0.21	2.15±0.11	0.31±0.18	00	6.41±0.14	2.07±0.18	0.19±0.07	00			
Liver	4.07±0.19	1.17 ± 0.2	0.25±0.15	00	4.12 ± 0.2	1.3 ± 0.10	0.10 ±0.06	00			

The obtained results revealed that florofenicol residues in liver and kidney in both healthy and diseased duckling treated with florofenicol were high at 1st day of clearance period and completely disappeared at 9th days of clearance period. High residue of florfenicol was found in kidney followed by liver. Florofenicol residues in liver and kidney disappeared at 8 day of clearance period (44, 45). Florofenicol residues in kidney more than in liver (46, 47).

Our findings revealed that duckling infected with Pasterulla multocida and treated with florfenicol showed disappearance of clinical signs, reduced mortality rate (5%), ameliorate the adverse effects and return leukogram and biochemical parameters to normal levels. This finding was previously recorded (26) which showed that pasteurellosis treated with florfenicol induce improvement of clinical signs and hematobiochemical parameters. This improvement may be due to antimicrobial effect of florfenicol (2).

From the obtained results in the current study, it could be concluded that florfenicol is effective against Pasterulla multocida in duckling

REFERENCES

- 1.Mohan K and Kumar P (2008): Pasteurellosis in a Duck Vet World 1(12) 48-56.
- 2.Ahmed Mona A, Fatma M Yousseff and Amina A (2012): Clinicopathological studies on bacterial respiratory diseases in ducks. Zag. Vet .J. 40(4) 46 -57.
- 3 Jonas M, Morishta T and Jhja J (2001): characterization of 8Pasterulla multocida isolates from avian cholera outbreak in Indonesia. Avian Dis.45:34-42.

- 4.Boglárka S, Krisztián B and Tibor M (2010): Characterization of the ptfA gene of avian Pasteurella milt strains by allelespecific polymerase chain reaction. J. of Vet. Diagnostic Investigation Vol. 22 Issue 4, 607-610.
- 5.Ganti A (2003): Avian Pasterullosis. In Textbook of Vet. Pathology, 7th ed. Satish Kumar Jain, CBS Publishers and Distributors, New Delhi, p. 612.
- 6.Baki A Dewan I and Mondal M (1993): pasteurellosis in duck. Agr 27(4) 33-34
- 7.Carter G and De Alws M (1989): Hemorrhagic septicaemia. In: Pasteurellosis, Adlam, C and Rutter, J (Ed) London: Academic Press, 13–160.
- 8.Kobal S (2004): Florfenicol and use in Vet. Med. Vet. Novice (30) 4, 12-15.
- **9.Sams R.** (1994): Florfenicol: chemistry and metabolism of a novel broad spectrum antibiotic.In: Proc. of VIII World Buiatrics Conf. Bolo, Italy, 13-17.
- 10.Cannon M, Harford S, Harford S and Davies J (1990): Acomparative study on the inhibitory action of chloramphenicol, thimphenicol and some fluorinated derivatives. J.Antimi. Chem.26:307-317.
- 11. Soback S, Paape M and Varma K (1995): Florfenicol pharmacokinetic in lactating cows after I/V, I/M and intramammary administrations. J. Vet. Pharma. Therap., 18 (6): 413-417.
- 12.Cruickshank R, Duguid J, Marmion B and Swain R (1975): Medical Microbiology.(2) Practice of Med. Microbiology.12thEd. Churchill Livingstone, London
- 13.Okerman L, Spanoghe L and Bruycker R (1979): experiental infection of mice with Pasterulla multocida strain isolated from rabbits. J. Comp. Path. 89:51-55.
- 14.Quinn P, Markery B, Carter M, Donnelly W and Leonard F (2002):

 Veterinary Microbiology and Microbial Diseases. Block well Sci Ltd.

- 15.Baron E Petrso L and fingld S (1992) Diagnostic Microbiology 8thEd Mosb17.
- 16 Jain N (2000): Schalm Veterinary Haematology, 8thEd Lea& Philadelphia.
- 17.Doumas B, Certor R, Peers T and Schafler R (1981): Acandidate refere-nce method for determination of total protein in serum. Clin. Chem. (27) 1642.
- 18.Henry R Cannon D and Winkelman J (1974): Clinical Chemistry: Principals and techniques p 437 440, Harper and Row, Hagerstown.
- 19.Reitman S and Frankel S (1957): Colorimetric determination of SGot and SGpt activity. Am. J. Clin. Path. (28) 56 – 59.
- 20 John D (1982): Clinical Lab. Method for Determination of Alkaline Phosphates 9th Ed. 580-581.
- 21.Coalombe J and Faurean l (1963): A new simple method for colorimetric determination of urea. Clin. Chem. (9)102-108.
- 22.Husdan H and Roporpot A (1968):
 Estimation of creatinine.Clin Chem
 (14)22 23. Arret B Johnson A and
 Kirshboum A (1971): outline for
 microbiological assay of antibiotics, 2nd
 revision. Pharma. Sci. 60:1689-1694.
- 24.Petrie A and Watson P (1999): Statistics for Veterinary and Animal Science 1st Ed. 90-99, the Blackwell Science LTd, United Kingdom.
- 25.Radad K and Fatma A Mostafa (2006):
 Studies on pasteurella milt and other bacterial pathogens associated with some problems in duck farms in Assuit Governorate. Assuit Vet. Med. J., 52 (108): 336-353.
- 26.Koshaik M, Abdel-Aziz S, Abdel-Alim Fand Kamel M (2008): Treatment of Pasteurellosis in Ducks. 9th Vet. Med. Zag. Conf. 861-872.
- 27. Hornish E and Susan F (2002): Cephalosporins in veterinary medicine:

- Ceftiofur use in food animals. Current Topics in Med. Chem. 2 (7): 717 731.
- 28. Abdel Aziz M (2002): Hand Book of Veterinary Pharmacology 5th Ed. Faculty of Vet. Med, Zag Uni.
- 29.El-Banna H. (1998): Pharmacokinetics of florfenicol in normal and Pasteurella infection Muscovy ducks. Brit. Poult. Sci. 33 (4): 492 496.
- 30.Abdel-Moneim A and El-Gendy A (1998):
 Comparative biochemical and haematological effects of florfenicol and chloramphenicol in albino rats. J. Egypt. Gen. Soc. Zool., 27 (A): 151-169
- 31.Swapnil S, Madhu S and Yamin V (2011):Effect of Mannan Oligosacch-aride on Pathology of Fowl Cholera in Broiler. Asian J. of Poul Sci, 5:86.
- 32. Zainab F (2006): Efficacy of fosfomycin and florfenicol in chickens. M Sc. Thesis Faculty of vet Medicine, Zagazig University.
- 33.Hend M (2007): Efficacy of some antimicrobials in chickens. M Sc. Thesis Fac. of vet Med., Zag. Uni.
- 34.Abdullah O, Tawfik M and Selim M (2001): Evaluation of concurrent use of ceftiofur sodium and isoflupredone acetate for treatment of pasteurello- sis in ducklings at Sharkia province. J. Egypt Vet. Med. Ass. 61(4): 321 337.
- 35. Abozed A Omima (1987): Aiver function tests in relation to some bacterial diseases in chicken.MVSc. Thesis Fac. of Vet. Med. Zag Uni.
- 36. Kaneko J (1980): Clinical Biochemistry of Domestic Animals. Academic Press. Inc, 4th Ed. New York, London, Tokyo. pp. 365 391.
- 37.Butler E (1983): "Plasma proteins." In: Physiology and Biochemistry of the Domestic Fowl, edited by Freeman. B. M. Academic Press.
- 38.Abd-Alla, A (1997): Clinicopathological studies on the effect of Pasteurella milt in

- chicken and ducks. Egypt. J Comp Path& Clin. Path 10 (2) 49-59.
- 39.Hamed E (2005):pharmacological studies on florfenicol and its interaction with streptomycin in chickens MVSc Thesis Faculty of Vet. Med. Zag. Uni.
- 40.Ali A and Youssef A (2003): Bacteriological studies and biochemical parameters of respiratory infection in ostriches.Vet. Med. J. Giza, 51(2)89–90.
- 41.Campbell T and Coles E (1986): Avian Clin. Pathol. In Vet. Clin. Pathol. 4thEd. W.B.Saunders Company Philadelphia, London and Toronto.
- 42.Halliwell W (1981): Serum chemistry profiles in health and disease of birds of prey. In Recent advances in the study of rapter disease, edited by cooper. J and Greenwood A Chiron Publication. Lit. West. Yorkshire England.

- 43.EL-Sayed E, EL-Azzawy H and Gheith S (2000): "Effect of ceftiofur sodium on ducks infected with Pasterulla multocida Egy. J. Agric. Res78 (1)
- 44.El- Banna H, Zaghlol A and Rehab M (2007): Efficacy and tissue reside depletion of florofenicol (water soluble formulation) in healthy and E. coli infected broiler chickens. Res. J. of Biological Sci. 2(3) 319-325.
- 45. Samah K, Eslam H and Ola H (2012): Residue Withdrawal of Florfenicol from Serum and Edible Tissues of Broiler Chickens J. of American Sci; 8(12).
- 46.Afifi N and El Sooud K (1997): concentration and pharmacokinetics of florofenicol in broiler chickens.Br. Poul. Sci., 38:425-428.

الملخص العربى دراسات بكتريولوجيه بيوكيميائيه على البط البيكينى المصاب معمليا بالباستريلا مع محاوله العلاج

د. حسام حسن علام على صلاح بيومى ، رضا حسن ذكى ، نجوى عباس بحر أ أقسام (الدواجن الكيمياء (٢-٦و٤)) معهد بحوث صحة الحيوان (فرع الزقازيق (٢-١) كفر الشيخ واسماعليه)

تم تجميع عدد ١٥٠ عينه من الأعضاء الداخلية لبط بيكيني عمر ١ وحتى شهر (٧٥ بطه سليمه ظاهريا, ٣٥ بطه يظهر عليها أعراض متمثله في الإسهال والكساح و ٤٠ بطه نافقه حديثا) الفحص البكتريولوجي. وبعد الفحص البكتريولوجي وجد ميكروب الباستريلا مالتوسيدا في عدد ٤٣ عينه (٢٨,٦٧%) من ١٥٠ عينه تم فحصها موزعه كالاتي (٥ بطه سليمة, ١٣ بطه مريضه و ٢٥ بطه نافقه. وبعمل اختبار الحساسية لتلك المعزولات وجد أن الفلور فينيكول أكثر المضادات الحيوية المستخدمة تأثيرا على البستريلا مالتوسيدا المعزوله عن باقي المضادات الحيوية المستخدمه.

في هذه الدراسة تم استخدم عدد ١٦٠ من البط البيكيني عمر يوم واحد (٨٠ بطه سليمة و ٨٠ بطه مصابه اصابه اصطناعية بالياستريلا مالتوسيدا. قسم البط إلى أربع مجموعات متساوية. المجموعة الأولى بطسليم ظاهريا واكلينكيا ولم يعالج (مجموعة ضابطة) والمجموعة الثانية بطسليم ظاهريا واكلينكيا وتم اعطاؤه الفلور وفينيكول بجرعة المحمم /كجم من وزن الجسم لمده ٥ أيام متتالية المجموعه الثالثة بط مصاب اصابه اصطناعية بالباستريلا مالتوسيدا ولم يتم علاجه إما المجموعه الرابعة بط مصاب اصابه اصطناعية بالباستريلا مالتوسيدا ويتم علاجه باستخدام الفلور وفينيكول بنفس الجرعة والمده السابقه. يتم دراسة تأثير الباستريلا على نسبه الوفيات ووزن الجسم. يتم اخذ عينات دم من البط في كل المجموعات وذلك لقياس بعض الوظائف المناعية والبيوكيميائيه. يتم ذبح عدد ٥ بطه عند فترات مختلفة إثناء وبعد نهاية العلاج ويتم اخذ عينات من الكبد والكلى لتعيين بقايا الفلور وفينيكول.

وأظهرت النتائج أن البط السليم والمعالج بالجرعة العلاجيه من الفلور وفينيكول أدى إلى حدوث زيادة معنوية في وزن الجسم, العدد الكلى لكرات الدم البيضاء, الخلايا الليمفاوية ونقص معنوي في خلايا الهيتروفيل بجانب زياده غير

معنويه في معدل التحويل الغذائي الخلايا الملتهمة الكبيرة, الخلايا القاعدية, الخلايا الحامضيه, البروتين الكلى, الزلال, النسبه بين الزلال والجلوبيولين, حمض اليوريك والكرياتينين AST, ALT, ALP . حمض اليوريك والكرياتينين

البط المصاب اصابه اصطناعية بالباستريلا مالتوسيدا ظهرت عليها أعراض مرضيه تتمثل في حدوث أصوات غير طبيعية والامتناع عن الأكل والإسهالات بجميع الألوان وارتفاع نسبة النافق ٣٠% و وجود زياده معنويه في عدد الخلايا الملتهمة الكبيرة الجلوبيولين الكلى والبيتا والجاما جلوبيولين, AST, ALP, حمض اليوريك والكرياتينين بجانب نقص معنوي وزن الجسم, خلايا الهيتيروفيل, الزلال إلفا حلوبيولين ,النسبه بين الزلال والجلوبيولين ونقص غير معنوي في معدل التحويل الغذائي العدد الكلى لكرات الدم البيضاء, الخلايا الليمفاويه, الخلايا الحامضيه, البروتين الكلى وزيادة غير معنوية في ALT

وقد دلت نتائج الدراسة على أن الفلوروفينيكول له بقايا في الأنسجة أثناء وبعد العلاج وكان أعلى منسوب لبقايا الفلوروفينيكول في الكلى يليها الكبد , ولكن الفلوروفينيكول اختفي من الأنسجة بعد مرور ٩ يوم من الحقن

من كل ما سبق نلاحظ أن أستخدام الفلور وفينيكول بالجرعة العلاجية لـة تأثير فعال في علاج الإصابة بالباستريلا وأدى إلى اختفاء الأعراض الظاهرية وأدي إلى عودة هذه الوظائف إلى المستوى الطبيعي.