Zagazig Veterinary Journal Volume 45, Number 1, p. 74-81, March, 2017 ©Faculty of Veterinary Medicine, Zagazig University, 44511, Egypt DOI: 10.21608/zviz.2017.7689.

Hormonal and Immunological responses to Coleus forskohlii treatment in Female Rats with Experimentally Polycystic Ovaries Syndrome

Hoda M.L. Abd-Allah^{1*}, Mona M. AbdAlhady², Magda N.A. Abdelall¹, Sahar N. Mohamedy¹, Waheeb D.M. Alharbi³ and Adil O. Bahathiq³

¹Animal Health Research Institute, Zagazig Laboratory, Egypt

²Physiology Department, Faculty of Veterinary Medicine, Zagazig University, 44511, Egypt ³Physiology Department, Faculty of Medicine, Umm al Qura University, Makkah, KSA

Article History: Received: 2/2/2017 Received in revised form: 5/3/2017 Accepted: 12/3/2017

Abstract

Polycystic ovaries syndrome (PCOS) is one of the most prevalent causes of reproductive failure. Its pathogenesis may be hormonal and immune disorder with hyperactivity of the sympathetic nervous system. The present study was conducted to demonstrate the effect of coleus forskohlii roots extract (CFE) in modulating the levels of some hormones and immune parameters of rats with androgen induced PCOS. Fifty immature female albino rats were divided into control group and androgen treated group that injected subcutaneously with 60 mg/kg BW/day androgen (Cidotestone) for 35 days to induce PCOS. Each group was subdivided into two subgroups. The first two subgroups received saline and 25 mg/kg BW of CFE, respectively while the other two subgroups received the same doses of Cidotestone and Cidotestone + CFE, respectively for 3 weeks. Blood samples were collected at the end of the experiment to estimate the levels of corticosterone, beta endorphin, total leucocytic count (TLC), cluster of differentiation 4 ($CD^{+}4$), cluster of differentiation 8 (CD⁺8), interleukin 2 (IL-2), interleukin 3 (IL-3), interleukin 4 (IL-4), immunoglobulin G (IgG) and immunoglobulin A (IgA). The results revealed a significant decrease in the concentrations of β endorphin, IL-2, TLC, CD⁺4, CD⁺8 and IgG, while corticosterone and IL-4 showed a significant increase in the androgen induced PCOS group. Rats treated with Cidotestone + CFE displayed normal values of all tested parameters except corticosterone, suggesting role of CFE in mediating both humoral and cellular immunity.

Keywords: Coleus forskohlii, Rats, Polycystic Ovaries Syndrome

Introduction

Polycystic ovary syndrome (PCOS) is a endocrinological complex disorder hyperandrogenism characterized by and ovulatory dysfunction [1] and its etiology remains obscure. PCOS assumed to be due to neuroendocrine defect leading to an and exaggerated LH pulse frequency amplitude, a deficiency in insulin action leading to hyperinsulinemia or ovarian changes in FSH response [2]. Interestingly there is a close relationship between the endocrine system, the sympathetic nervous system and the immune system [3]. Basic symptoms of PCOS such as anovulation and follicular cysts were produced in female mice by injection of estrogen, testosterone or cortisone during immune adaptive period resulting in interference with the final stage of thymus gland development that alters the evolution of self versus non-self recognition Additionally, Öner and Ozan [4,5]. [6]

suggested that steroids forestall the production of regulatory T cells.

Rats with estradiol valerate induced PCOS have reduced hypothalamic β-endorphin concentration and an increase of µ-opioid binding reflecting a chronic up-regulation of the receptor in response to compromised β endorphin input [7]. Leukocytes can express opioid receptors [8] and under certain circumstances also synthesize and release βendorphin themselves providing the molecular mechanisms for communication with the neuroendocrine system [9]. It is documented that women with PCOS have decreased frequencies of circulating CD⁺8T cells and natural killer (NK) cells [10], altered cytokine responses and an increased number of activated T cells in follicular fluid [11,12]. The cyclic adenosine monophosphate (cAMP) can differentially regulate multiple cell processes and act as a positive regulator of a number of

74

immune function genes [13], through different pathways; including cyclic nucleotide gated ion channels, cAMP-activated protein kinases, or exchange proteins directly activated by cAMP [14,15]. Since forskolin is one of cAMP generator and represents as an active constituent of coleus forskohlii plant [16]. Its role as aromatase-agonist-like drug that can directly induce follicular development and shorten the treatment course of PCOS women, irrespective of the hyperandrogen state [17]. Hence, the present study was conducted to determine if rats with androgen- induced PCOS have alterated carticosterone, ßendrophine and immune response. The possible role of coleus forskorskohlii roots extract in alleviation these effects were also studied.

Material and methods

Animals and experimental design

Fifty pre-pubertal female albino rats (23 days old) were obtained from laboratory animal unit Faculty of Veterinary Medicine, Zagazig University and kept at a temperature of 25±2°C with a 12 light: 12 dark cycle. All animals were freely accessed to pelleted food and water and kept for one week before the beginning of the experiment. Animals were divided into two equal groups: the first group was injected with saline subcutaneously (S.C) and the second group was injected with (androgen cidotestone synthetic steroid, Chemical Industries Development Company, Giza, A.R.E) 60 mg/kg BW/day for 35 days for inducing PCOS [18] that was verified by vaginal smear analysis and histopathological examination. After verifying the induction of PCOS, the first group was subdivided into two equal subgroups, the first subgroup was left as control group (saline S.C and per os (PO)) and the second subgroup was given CFE, 25mg/kg BW PO (Ethanolic extract of coleus forskohlii roots that prepared according to Farias et al. [19] and saline S.C. The second main group was subdivided into two equal subgroups, the first subgroup injected with cidotestone 60 mg/kg BW/day S.C + saline PO and the second subgroup was injected with cidotestone + CFE treatment group (with the same previous doses of cidotestone and CFE). All treatments were given daily for 21 consecutive days. Rats were fasted for 6 hours and then ten rats from each group were sacrificed and blood was collected in ice chilled heparinized tubes for hormones and immune analyses. Blood samples were centrifuged at 3000 rpm for 15min and plasma was separated, frozen and stored at 20°C until assay.

Biochemical analysis

Hormonal and some immune parameters were estimated by enzyme-linked immunesorbent assay (ELISA) kits to quantify the plasma beta-endorphin (ELISA kit, cat. No. MBS 704725), coticosterone (ELISA kit, cat No. MBS 705109), cluster of differentiation 4 (CD⁺4, ELISA kit, cat. No MBS 263383), cluster of differentiation 8 (CD⁺ 8, ELISA kit, cat. No. MBS 2503027), interleukin 2 (ELISA kit, cat. No. MBS 723159), interleukin 3 (ELISA kit, cat. No MBS 730102), interleukin 4 (ELISA kit, cat. No MBS 175989), immunoglobulin A (ELLSA kit, cat. No. MBS2500774) and immunoglobulin G (ELISA kit, cat. No. MBS 261432). Blood was used to determine of leucocytic count (TLC) using a hematological analyzer (coulter Micro Diff II Coulter Electronics Ltd, USA).

Statistical analysis

All statistics for the present data were determined using ANOVA (F-Test) followed by LSD (Least Significant Difference) for comparative of means using SPSS version 14 [20].

 Table 1: Effect of coleus forskohlii root extract (CFE) on corticosterone and beta endorphin in rats with androgen induced polycystic ovaries

Parameters	Treatments				
	Control	Androgen	Androgen+CFE ¹	CFE ¹	
Corticosterone(ng/ml)	0.63 ± 0.022^{d}	$1.01 \pm 0.047^{\circ}$	1.37±0.099 ^b	1.99±0.062 ^a	
Endrophine (pg/ml)	68.79 ± 2.85^{b}	54.93±1.57°	61.29±1.99 ^c	89.79±2.85 ^a	

Data represented as mean \pm SE (standard error) and the different letter in the same row were statistically significant at (p \leq 0.05) using LSD (least significant difference). ¹CFE= Coleus forskohlii roots extract.

Results

The levels of corticosterone hormone and beta endorphin in androgen induced PCOS and after treatment with CFE were presented in Table 1. It was clear that corticosterone levels increased significantly both with androgen induced PCOS and after treatment with CFE in comparison with the control group. Whereas the levels of beta endorphin exhibited a significant decrease in androgen induced PCOS group compared with the control group. In contrast, their concentrations were elevated in androgen administrations with CFE or CFE alone when compared with the control group. The impact of androgen induced PCOS on total leucocytic count and the recurrences of CD4 and CD8T cells were shown in Table 2. All these parameters were significantly lower in rats with PCOS group when compared with the control group. However, CFE treatment could restore these parameters to their control values.

Table 3 revealed a significant decrease in the concentration of IL2 with a significant increase of IL4 in androgen induced PCOS group, while IL3 concentration did not differ in this group compared with the control group. Treatment with CFE abolished alteration in cytokines responses. The results revealed a significant decrease in the concentration of IgG in the hyperandrogenized group, while IgA concentration displayed no significant response within this group compared to the control. Treatment with CFE evoked a significant elevation in IgG concentrations.

Table 2: Effect of coleus forskohlii root extract (CFE) on total leucocytic count, CD+4 and CD+8 in rats with androgen induced polycystic ovaries

Parameters	Treatments				
	Control	Androgen	Androgen+CFE ¹	CFE ¹	
² TLC X 10 ³	8.75±0.52 ^b	$6.34 \pm 0.70^{\circ}$	7.89 ± 0.60^{bc}	10.76±0.25 ^a	
3 CD4 (ng/mL)	$0.51 \pm 0.022^{\circ}$	0.32 ± 0.011^{d}	0.78 ± 0.043^{b}	1.02 ± 0.50^{a}	
⁴ CD8(pg/mL)	18.7±0.54 ^b	13.10±0.38 ^c	25.78±1.39 ^a	28.10 ± 0.62^{a}	

Data represented as mean \pm SE (standard error) and the different letter in the same row were statistically significant at (p \leq 0.05) using LSD (least significant difference).¹CFE= Coleus forskohlii roots extract. ²TLC= total leucocytic count. ³CD4=cluster of differentiation 4. ⁴CD8= cluster of differentiation 8.

Discussion

There is a possible link between ovarian functionality and immune response during cytogenesis induced by steroid hormones. The present data revealed a significant increase in the levels of corticosterone in both androgen induced PCOS and androgen+ CFE groups, while there was a significant decrease in the plasma level of β endorphin and recurrences of circulating CD⁺4 and CD⁺8T cells in the hyperandrogenized group when compared with the control group. Coleus forskohlii extract treatment elevated β endorphin and the T lymphocyte activity in hyperandrogenized rats to simulate the control values.

Glucocorticoid prereceptor metabolism enhanced in 5α -dihydroxytestosterone treated rats (a model of PCOS) that assessed by elevated intracellular corticosterone [21]. On the other hand, Keefe *et al.* [22] showed no differences in serum concentration of the

dihydroepiandrosterdione. adrenal steroids cortisol, corticosterone and their 11-deoxy precursors in woman with PCOS suggested an increase in their production and clearance [23]. In the present study, elevated levels of corticosterone in hyperandrogenized rats treated with CFE may be due to forskolininduced nuclear accumulation of the recognized corticotropin-releasing hormones transcriptional regulators [24] and enhanced the normal glucocorticoid receptor function producing adrenal glucocorticoid that is essential for homeostasis and survival during severe stress situation [24,25]. In the current β endorphin concentration study, was decreased in rats with androgen induced PCOS. Similar results were obtained by Lobo et al. [26] in human with PCOS and in rats with estradiol valerate induced PCOS [27], indicating upset β endorphin production with this disease, since β endorphin aroused a tonic inhibitory control of gonadotrophin hormones

pulse generator [28], leading to the suppression of plasma LH pattern that is characterized the steroid–induced PCOS model [7]. The cAMP is considered an important second messenger system in regulation of hormone secretion and proopiomelanocortin [29] as well as having a critical role in the differentiation of β endorphin neurons [30]. This may explain the normalization of β endorphin concentration after CFE (cAMP stimulating agent) administration.

Table 3: Effect of coleus forskohlii root extract (CFE) on interleukins and immunoglobulins in rats with androgen induced polycystic ovaries

Parameters	Treatments				
	Control	Androgen	Androgen+CFE ¹	CFE ¹	
2 IL2 (pg/mL)	211.8±6.92 ^a	152.69±1.45 ^c	195.4±1.63 ^b	212 ± 4.84^{a}	
$^{3}IL3 (pg/mL)$	237.6±4.75	240±4.93	236±4.16	237.4±4.97	
4 IL4 (pg/mL)	137.8 ± 1.68^{b}	143.61±1.18 ^a	134.77 ± 1.49^{b}	$114 \pm 0.92^{\circ}$	
IgG (ng/mL)	3.81 ± 0.12^{b}	2.71±0.180 ^c	5.16±0.024 ^a	5.44±0.133 ^a	
IgA (ng/mL)	41.38±0.921	42.70±1.53	38.28±1.41	38.02±0.904	

Data represented as mean \pm SE (standard error) and the different letter in the same row were statistically significant at (p \leq 0.05) using LSD (least significant difference).¹CFE= Coleus forskohlii roots extract. ²IL2=interleukin 2. ³IL3=interleukin 3. ⁴IgG= immuoglobin G. ⁵IgA= immuoglobin A.

Androgen (Testosterone, dehydroepiandrostrone or androstenedione) structure is determinant in either inhibiting or enhancing T lymphocyte proliferation, but testosterone was more potent than dehydroepiandrosterone in suppression of the thymocyte proliferation [31]. The present findings of significantly decreased total leukocytic count, frequencies of circulating CD⁺4 and CD⁺8 cells in hyperandrogenized rats, were in agreement with the previous reported results [27,31,32], suggesting an induction of oxidative stress caused by androgen that resulted in an increase of the nitric oxide synthase and nitric oxide, consequently lead to inhibition of the T cell proliferation. Additionally, steroids may forestall the production of regulatory T cells [5]. Treatment hyperandrogenized rats with CFE may be resulted in regulation of reactive oxygen species and the induction of AMP activated protein kinase (AMPK) of T lymphocytes [33,34].

The present study revealed a decreased IL-2 and IgG concentration in rats with androgen induced PCOS that was restored to control levels after CFE treatment. This result was consistent with the previous studies that have shown that dehydro-epiandrosterone (a week androgenic steroid) significantly reduced the Th₁ cytokines IL-2 and had no effect on the production of the Th₂ cytokine IL-4 in vitro after stimulation with the mitogens conconavalin A [35]. However, Meikle et al. [36] indicated that androgen is a steroid hormone that is directly involved in the regulation of IL-2 production by both normal and some T cell hybridomas. Th₁ (make IL-2 and IFN-y) and Th₂ (make IL4, IL5 and IL6) cells are using different transmission pathways after T cell receptor (TCR) mediated stimulation. The protein kinase C pathway is the major system of activation in Th₁ cells, while different second messengers are generated in Th₂ cells after activation [37]. The increased IL-2 and IgG in CFE treated hyperandrogenized rats in the present study may be explained an increase of opioid peptides such as β -endorphin that affect both free intra-cellular calcium and intracellular cAMP concentrations [38]. The rise in intracellular calcium is dependent on calcium released from intracellular stores [39], in response to the generation of inositol 1,4.5triphasphate [40] and the influx of extracellular calcium [39]. Interestingly, forskolin could increase calcium influx through voltage-gated channels [41]. Intracellular calcium mobilization is an important early event involved in T cell activation and proliferation [42] and both types of T cell were able to stimulate B cells to secrete antibodies like IgG [43]. Paradoxically, Muñoz et al., [37] recorded that forskolin

inhibited T cell receptor-mediated IL-2 production and proliferation in Th_1 cells suggested their differential sensitivity to high levels of cAMP

Conclusion

The present data revealed that opioid and immune systems were impaired in hyperandrogenized rats (PCOS model) and the CFE treatment could restore most of these functions indicating its essential role in modulating both humoral and cellular immunity.

Conflict of interest

The authors have no conflict of interest to declare.

References

- Yen, S.S.C. (1999): Polycystic ovary syndrome (hyperandrogenic chronic anovulation). In:Yen SSC, Jaffe, RB (eds.), Reproductive endocrinolo gy, 4th^{ed}. Philadelphia: WB Saunders Co.; 436-476.
- [2] Matalliotakis ,I.; Kourtis, A.; Koukoura ,O. and Panidis, D. (2006): Polycystic ovary syndrome: etiology and pathogenesis. Arch Gynecol Obstet, 274(4):187-97.
- [3] Chrousos, G.P. and Gold, P.W. (1992): The concepts of stress and stress system disorders. JAMA, 267(9):1244-1252.
- [4] Chapman, J.C.; Min, S.; Kunaporn S.; Tung, K.; Shah, S. and Michael, S.D. (2002): The administration of cortisone to female B6A mice during their immune adaptive period causes anovulation and the formation of ovarian cysts. Am J Reprod Immunol,48(3):184-189.
- [5] West, L .J. (2002): Defining critical windows in the development of the human immune system. Hum Exp Toxicol, 21(9-10):499-505
- [6] Öner, H. and Ozan, E. (2002): Effects of gonadal hormones on thymus gland after bilateral ovariectomy and orchidectomy in rats. Arch Androl, 48(2):115-126.
- [7] Desjardins, G.C.; Beaudet, A. and Brawer, J.R. (1990): Alterations in opioid parameters in the hypothalamus of rats with

estradiol-induced polycystic ovarian disease. Endocrinol,127(6):2969-2976.

- [8] Sharp, B.M.; McAllen, K.; Gekker, G.; Shahabi, N.A. and Peterson, P.K. (2001): Immunofluorescence detection of delta opioid receptors (DOR) on human peripheral blood CD4T cells and DORdependent suppression of HIV-1 expression. J Immunol, 167(2):1097-1102
- [9] Sharp, B. and Linner, K. (1993)::What do we know about the expression of proopiomelanocortin transcripts and related peptides in lymphoid tissue? Endocrinol, 133(5):1921A-1921B.
- [10] Turi, A.; Di Prospero, F.; Mazzarini, A.; Costa, M.; Cignitti ,M.; Garzetti ,G.G. and Romanini, C. (1988): Lymphocytes subset in hyperandrogenic women with polycystic ovarian disease. Acta Eur Fertil, 19(3):155-157.
- [11] Amato, G.; Conte, M.; Mazziotti, G.; Lalli, E.; Vitolo, G.; Tucker, A.T.; Bellastella, A.; Carella, C. and Izzo, A. (2003) : Serum and follicular fluid cytokines in polycystic ovary syndrome during stimulated cycles. Obstet Gynecol, 101(6):1177-1182.
- [12] Gallinelli, A.; Ciaccio, I.; Gianella, L.; Salvatori, M.; Marsella, T. and Volpe, A. (2003): Correlations between concentrations of interleukin-12 and interleukin-13and lymphocyte subset in the follicular fluid of women with and without polycystic ovary syndrome. Fertil Steril, 79(6):1365-1371.
- [13] Hertz, A.L.; Bender, A.T.; Smith, K.C.; Gilchrist, M.; Amieux, P.S.; Aderem, A. and Beavo, J.A. (2009): Elevated cyclic AMP and PDE4 inhibition induce chemokine expression in human monocyte-derived macrophages. Proc Natl Acad Sci, 106(51):21978-21983.
- [14] de Rooij, J.; Zwartkruis, F.J.; Verheijen, M.H.; Cool, R.H.; Nijman, S.M.; Wittinghofer, A. and Bos, J.L. (1998): Epac is a Rap1 guanine-nucleotideexchange factor directlyactivated by cyclic AMP. Nature, 396(6710):474-477.

- [15] Kawasaki, H.; Springett, G.M.; Mochizuki, N.; Toki, S.; Nakaya, M.; Matsuda, M.; Housman, D.E. and Graybiel, A.M. (1998): A family of cAMP-binding proteins that directly activate Rap1.Science, 282(5397):2275-2279.
- [16] Seamon, K.B.; Padgett, W. and Daly, J.W. (1981): Forsfolin: unique diterpene activator of adenylatecyclase in membranes and indirect cells. Proc Natl acad Sci, 78(6):3363-3367.
- [17] Chen, J.; Shen, S.; Tan, Y.; Xia, D.; Xia, Y.; Cao, Y.; Wang, W.; Wu, X.; Wang, H.; Yi L.; Gao, Q. and Wang, Y. (2015): The correlation of aromatase activity and obesity in women with or without polycystic ovary syndrome. J Ovarian Res, 8(1):11
- [18] Jang, M.; Lee, M.J.; Lee, J.M.; Bae, C.S.; Kim, S.H.; Ryu, J.H. and Cho, I.H. (2014): Oriental medicine Kyung-Ok-Ko prevents and alleviates dehydroepiandrosterone-induced polycystic ovarian syndrome in rats. PLoSOne, 9(2):1-14.
- [19] Farias, F.D.; Souza, T.M.; Viana, M.P.; Soares, B.M.; Cunha, A.P.; Vasconcellos, I.M.; Ricardo, N.M.P.S; Ferreira, P.M.P.; Melo, V.M.M. and Carvalho, A.F.V. (2013): Antibacterial, antioxidant and anticholin- esterase activities of plant seed extract from Brazilian semiarid region . BioMed Research Int, 2013:1-9.
- [20] SPSS 14 (2006)."Statistical Package for SocialScience, SPSS for windows Release14,0.0, 12 June,2006" Standard Version,Copyright SPSS Inc., 1989-2006, All RightReserved Copyright® SPSS Inc.
- [21] Nikolić, M.; Macut, D.; Djordjevic, A.; Veličković, N.; Nestorović, N.; Bursać, B.; Antić, I.B.; Macut, J.B.; Matić, G. and Vojnović Milutinović, D. (2015): Possible involvement of glucocorticoids in 5α -dihydrotestosterone-induced PCOS-like metabolic disturbances in the rat visceral adipose tissue. Mol Cell Endocrinol, 399:22-31.

- [22] Keefe, C.C.; Goldman, M.M.; Zhang, K.; Clarke, N.; Reitz, R.E. and Welt, C.K. (2014): Simultaneous measurement of thirteen steroid hormones in women with polycystic ovary syndrome and control women using liquid chromatographytandem mass spectrometry. PLoS One, 9(4):1-7.
- [23] Rodin, A.; Thakkar, H.; Taylor, N. and Clayton, R. (1994): Hyperandrogenism in polycystic ovary syndrome. Evidence of dysregulation of 11 betahydroxysteroid dehydrogenase. New Engl J Med, 330(7):460-465.
- [24] Evans, A.N.; Liu, Y.; Macgregor, R.; Huang, V. and Aguilera, G. (2013): Regulation of hypothalamic corticotropin-releasing hormone transcription by elevated glucocorticoids. Mol Endocrinol, 27(11):1796-1807.
- [25] Eickelberg, O.; Roth, M.; Lorx, R.; Bruce, V.; Rudiger, J.; Johnson, M. and Block, L.H. (1999): Ligand-ind ependentactivation of the glucocorticoid receptor by beta2-adrenergic receptor agonists in primary humanlung fibroblasts and vascular smooth muscle cells. J Biol Chem, 274(2):1005-1010.
- [26] Lobo, R.A.; Granger, R.; Paul, W.L.; Goebelsmann, U. and Mishell, D.R. (1983): Psychological stress and increases in urinary norepinephrine metabolites, platelet serotonin, and adrenal androgens in women with polycystic ovary syndrome. Am J Obstet Gynecol, 145(4):496-503
- [27] Stener-Victorin, E. and Lindholm, C. (2004): Immunity and beta-endorphin concentrations in hypothalamus and plasma in rats with steroid-induced polycystic ovaries: effect of lowfrequency electroacupuncture. Biol Reprod, 70(2):329-333.
- [28] Wildt, L.; Sir-Petermann, T.; Leyendecker, G.; Waibel-Treber, S. and Rabenbauer, B. (1993): Opiate antagonist treatment of ovarian failure. Hum Reprod, 8(Suppl 2): 168-174.
- [29] Lundblad, J.R. and Roberts, J.L. (1988): Regulation of proopiomelanocortin gene

expression in pituitary. Endocr Rev, 9(1):135-158.

- [30] Sarkar, D.K.; Boyadjieva, N.I.; Chen, C.P.; Ortigüela, M.; Reuhl, K.; Clement, E.M.; Kuhn, P. and Marano, J. (2008):Cyclic adenosine monophosphate differentiated beta-endorphin neurons promote immune function and prevent prostate cancer growth. Proc Natl Acad Sci, 105(26):9105-9110.
- [31] Yao, G. and Shang, X.J. (2005): A comparison of modulation of proliferation of thymocyte by testosterone, dehydroisoandrosterone and androstenedione in vitro. Arch Androl, 51(4):257-265.
- [32] Solano, M.E.; Sander, V.; Wald, M.R. and Motta, A.B. (2008): Dehydroepiandrosterone and metformin regulate proliferation of murine T lymphocytes. Clin Exp Immunol, 153(2): 289-296.
- [33] Khatun, S.; Chatterje, N.C. and CaKilicoglu, U. (2011): Antioxidant activity of the medicinal plant Coleus forskohlii Briq. Afr J Biotechnol, 10 (13): 2530-2530.
- [34] Sander, V.; Luchetti, C.G.; Solano, M.E.; Elia, E.; Di Girolamo, G.; Gonzalez, C. and Motta A.B. (2006): Role of the N, N'-dimethylbiguanide metformin in the treatment of female prepuberal BALB/c mice hyperandrogenized with dehydroepiandrosterone. Reproduction, 131(3):591-602.
- [35] Powell, J.M. and Sonnenfeld, G. (2006): The effects of dehydroepiandrosterone (DHEA) on in vitro spleen cell proliferation and cytokine production. J Int Cytokine Res, 26(1):34-39.
- [36] Meikle, A.W.; Dorchuck, R.W.; Araneo, B.A.; Stringham, J.D.; Evans, T.G.; Spruance, S.L. and Daynes, R.A. (1992): The presence of a

dehydroepiandrosterone-specific receptor binding complex in murine T cells. J Steroid Biochem Mol Biol, 42(3-4):293-304.

- [37] Muñoz, E.; Zubiaga, A.M.; Merrow, M.; Sauter, N.P. and Huber, B.T. (1990): Cholera toxin discriminates between T helper 1 and 2 cells in T cell receptormediated activation: role of cAMP in T cell proliferation. J Exp Med, 172(1):95-103
- [38] Fowler, C.J. and Fraser, G.L. (1994): μ -, δ -, κ -opioid receptors and their subtypes. A critical review with emphasis on radiology and binding experiments. Neurochem Int, 24(5):401-426.
- [39] Ledbetter, J.A.; June, C.H.; Grosmaire, L.S. and Rabinovitch, P.S. (1987): Crosslinking of surface antigens causes mobilization of intracellular ionized calcium in T lymphocytes. Proc Natl Acad Sci, 84(5):1384-1388.
- [40] Guan, L.; Townsend, E.T.K.; Adler, M.W. and Rogers, T.J. (1994): Both T cells and macrophages are targets of kappa-opioid-induced immunosuppression. Brain Behav Immun, 8(3);229-240.
- [41] Jin, W.; Lee, N.M.; Loh, H.H. and Thayer, S.A. (1992): Dual excitatory and inhibitory effects of opioids on intracellular calcium in neuroblastoma x glioma hybrid NG108-15 cells. Mol Pharmacol, 42(6):1083-1089.
- [42] Tsien, R.Y.; Pozzan, T. and Rink, T.J. (1982): T-cell mitogens cause early changes in cytoplasmic free Ca2+ and membrane potential in lymphocytes. Nature, 295(5844):68-70
- [43] Snapper, C.M. and Paul, W.E. (1987): Interferon-gamma and B cell stimulatory factor-1 reciprocally regulate Ig isotype production. Science, 236(4804):944-947.

الملخص العربى

الاستجابة الهرمونية والمناعية للعلاج بنبات القسط في انا ث الجرذان المحدث بها تجريبا متلازمة تكيس المبايض هدى محمد لطفى عبدالله' ,منى محمد عبد الهادى' ,ماجدة نعمت عبدالحميد عبدالعال', سحر نصر محمد 'ى, وهيب دخيل الله محد الحريى', عادل عمر باحاذق' معهد بحوث حمة الحيوان- معمل الزقازيق أقسم علم وظائف الاعضاء-كلية الطب-جامعة ام القرى-مكة المكرمة-المملكة العربية السعودية

متلازمة تكيس المبايض (PCOS) هي واحدة من أكثر الأسباب شيوعا لضعف التناسل وقد يحدث هذا المرض نتيجة إضطرابات في الهرمونات أوفى جهاز المناعة مع النشاط المفرط في الجهاز العصبي الودي. وقد أجريت هذه الداسة لبيان تاثير مستخلص جذو□ نبات القسط في تعديل مستويات بعض الهرمونات وبعض المؤشرات المناعية في الجرذان المعالجة بهرمون الأند□وجين لإحداث متلازمة تكيس المبايض. تم تقسيم عدد خمسون من إناث الجرذان غير الناضجة أولا إلى مجموعتين: مجموعة ضابطة ومجموعة حقنت تحت الجلد بهرمون الأناحوجين (سيدوتيستون) بجرعة٦٠ملجم/كليو جرام من وزن الجسم لمدة ٣٥ يوم لإحداث متلازمة تكيس المبايض. ثم تم تقسيم كل من المجموعتين إلى مجموعاتان فرعيتان. المجموعاتان الفرعيتان الاوليتان (أعطيت محلول ملحي وأعطيت ٢٥ملجم/كجم من وزن الجسم من مستخلص جذو□ نبات القسط) والمجموعاتان الفرعيتان الأخريتان (أعطيت سيدوتيستون و سيدوتيستون + مستخلص جذو_ نبات القسط بنفس الجرعات السابقة) لمدة ٣ أسابيع. ثم تم تجميع عينات الدم عند نهاية التجربة وتم تقدير مستويات هرمون كو□تيكستو□ن والبيتا إندو∟فين والمجموع الكلي لخلايا الدم البيضاء ومجموعات التمايز (CD ⁺ 4)و(C⁺ 8) وانترلوكين ۲ وانترلوكين ۳ وانترلوكين ٤ والأجسام المناعية G وA وقدأظهرت النتائج إنخفاض ملحوظ في تركيز البيتا إندو∟فين والمجموع الكلي لخلايا الدم البيضاء ومجموعات التمايز (CD ⁺ 4) و(CD ⁺ 8) وانترلوكين ۲ والأجسام المناعية G بينما زاد مستويات هرمون كو_تيكستر∟ن وانترلوكين٤ في جرذان متلازمة تكيس المبايض المعالجة بهرمون الأندو_جين. في حين اظهرت جرذان متلازمة تكيس المبايض المعالجة بهرمون الأندو□جين + مستخلص جذو□ نبات القسط قيم طبيعية لمعظم المؤشرات ماعدا مستوى هرمون كو□تيكستو□ن مما يشير إلى دو□ مستخلص جذو□ نبات القسط في تعديل مستويات المناعةمما يساعد في تقصير مدة علاج متلازمة تكيس المبايض.