

Review Article

Sources and Toxicological impacts of Surface Water Pollution on Fish in Egypt

Amany T. Mohammed*, Magdy F. Abo-El fotoh, Mayada R. Farag,

Yasmina M. Abd El-Hakim and Walaa M. El Hady

Forensic Medicine and Toxicology Department, Faculty of Veterinary Medicine, Zagazig University, 44511, Egypt

Article History: Received: 6/11/2018 Received in revised form: 6/2/2019 Accepted: 23/2/2019

Abstract

Egypt has been listed as one of the most popular nations which are menaced through lack of water by the year 2025 due to overpopulation. The great deposition of highly polluted domestic and industrial effluents into its water ways, made a continuous and prompt damage to its surface and groundwater. The River Nile represents about 97% of Egypt's water resources; however winter rain and nonrenewable groundwater aquifers are also comprised. In Egypt, the main source of pollution is industrial wastes that are poured directly into the main water sources or through the municipal system. On the other hand, the pollution of the Mediterranean coast of Alexandria is mainly due to the discharge of wastes of industrial plants into the sea via Lake Marriott. The provincial populations have little or even no entry to drainage systems or wastewater treatment facilities and they rely only on the elimination of wastewater at the site of their production or collection. In the Delta region, drainage water is reused for irrigation after mixing with Nile water, while in Upper Egypt drainage water is disposed into the River Nile. Fertilizers and pesticides have been used on a wide range after the constructions of High Dam which resulted in weed flourishing which blocks the waterways, and provides habitats for Bilharzia snails. Egyptian fish aquaria exposed to toxicity due to water pollution from chemical plants and sewage pipes. Heavy metal pollution of Egyptian water is primarily produced by industrial and agricultural discharges. Other sources also include: coal and oil combustion, phosphate fertilizers, plastics and pesticides. Recently, the consumption of contaminated fish has been the possible cause of heavy metal poisoning in human. The aim of this review is to disclose the fundamental origin of water pollution in Egypt, and to elucidate the side effects of this pollution on fish.

Key words: Water pollution, Pesticides, Heavy metals, Fish, Egyptian water resources.

Introduction

Water is the key component to life existence as well as sustained nations and societies, as all activities of human beings depend on water, such as agriculture, industry, domestic and recreational activities, etc. [1]. Unfortunately, it is proposed that by 2025, 2.7 billion humans will be facing water scarcity globally [2], which might be a threat to the existence of life on earth. This subsequently renders the conservation of water and elimination of water polluting sources on a priority basis. The requirements of water have been increased owing to the industrialization and expanding populations. The additional supply is commonly fulfilled by the well water

or nearest available surface water. It may lead to incomplete treatment and substandard supply of drinking water [3].

Water pollution refers to changing of the chemical and physical characteristics of water from a beneficial state to one that is hazardous to organisms depending on water for their well-being [4]. Globally, industrial waste water represents the principal source of water pollution. The Uprising increase in modern industries, agriculture urbanization, tourism and human activities are the main sources for chemical pollution to both, aquatic environment and its coexisting ecosystems [5]. The developing countries are facing severe

*Corresponding author e-mail: (Amany_tharwat@yahoo.com), Forensic Medicine and Toxicology Department, Faculty of Veterinary Medicine, Zagazig University, 44511, Egypt.

environmental deterioration which continuously polluting water [6]. This might be due to natural activities including floods, sliding of mud, or volcanoes, but most of the activities contributing to water pollution are anthropogenic in nature, rendering water unusable [7].

Water pollution in Egypt is mainly due to the chemical and industrial processes. Because of discharging the industrial effluents and animal wastes into streams and rivers by factories and manufacturers in Egypt, several environmental and health problems were developed. Heavy metals have great awareness due to their accumulation, toxicity [8] and bio magnification [9] in the aquatic ecosystems.

Water pollution is categorized into a wide range of sources, including pesticides, organic waste products, fertilizers, heavy metals, and habitat modification.

1. Pesticides

1.1 Pesticides and water pollution

The term pesticide is a complex expression used for those chemicals used to resist, mitigate, deter, repel, prevent and destroy pests [10-11]. Air, water, and soil are contaminated with pesticides which may be due to its application in all the three media such as their direct application for controlling vectors, agricultural weeds, as well as aquatic flora. The accumulation of pesticides in soil and water is due to their escape with drainage, leakage via soil and transportation via air [12-14].

Pesticides cause several impacts of on water and those are related to the main ingredient and the existed impurities in the pesticide formulation. Furthermore, the added ingredients which are mixed with the main principal chemical compound (extenders, wetting agents, preservatives, diluents, emulsifiers, buffers, or solvents and adhesives), that is formed during microbial or photochemical degradation of the active ingredient [4].

1.2 Negative impacts of water pollution by pesticides creates negative impacts

1.2.1 Residues of pesticides in water

Pesticide residues are believed to be the key component of pollution [15]. All over the world, many types of pesticides are employed to cope with various pests and are designed in different ratios as herbicides (15%), fungicides (1.46%) and insecticides (80%) [11].

Pesticides lead to water bodies mainly through sediments (inorganic substrate), Algae, hydrophytes, and vascular branches or litter and moss (organic substances) [14- 16]. Pesticides can be cycled among different media such as water, non-target plants, and animals, wetlands, atmosphere and soil [17-19]. In water bodies, pesticides are found in higher concentration in stationary water while in sediments with negligible levels. The pesticides present in the water bodies pose a serious threat to the lives of aquatic organisms, as well as alter their regular life activities such as swimming, fitness, and species to species interaction [20].

Organochlorine compounds (OCPs) were found in high concentrations in about 23 sediment specimens gathered from Alexandria Harbor, Egypt as persistent concentrations. The authors found that polychlorinated biphenyls (PCBs) concentrations ranged from 0.9 to 1210 ng/g with 4 to 7 Cl-substituted biphenyls being the most prevalent PCBs congeners [21].

In a study about the levels of Organochlorine pesticides (OCPs) and polychlorinated biphenyls (PCBs) samples that were estimated in sediment gathered from 34 locations in Lake Qarun, Egypt. The study concluded that γ -HCH, endrin and chlordanes would be more concerned OCP species for the ecotoxicological risk in Lake Qarun [22].

Many authors estimated the distribution pattern of organochlorine residues in the Mediterranean Sea water (Egypt). They found that the concentration of total OCPs in surface sediments ranged from 0.11 to 50.73 with a mean of 14.52 ng/g dry weight [23].

In a recent study, it was found that HCHs, cyclodienes, DDTs, PCB concentrations in sediments of the Red Sea coast, Egypt were much lower than those recorded by the international organization [24].

1.2.2. Impacts of pesticides on marine life

1.2.2. a. Impacts on fish behavior

The survival of fish communities is indirectly affected by altering the plankton communities, which is a food source for different species of fish [25]. The presence of pesticides in water bodies altered the fish behavior [26]. Pesticides render the fish habitat unsuitable and adversely affect their escape behavior and locomotion, hence expose them to predators [27]. Pesticides also affect the predations/competition and life history of different fish taxa [28-29]. These indirect effects are considered as more severe and dangerous for the fish population than direct ones [30].

Sub-lethal concentrations of pesticides lead to diverse effects in fish. These changes may be in the behavior of the fish like disrupted schooling behavior, equilibrium loss, jerky movements, color change, erratic swimming, adapting vertical position, bottom sinking, jumping, motionlessness, sluggishness, hyper or hypo-excitability and increase in opercular beats, etc. These behavioral changes are observed in different fish species including *Cirrhinus mrigala*, *Channapunctatus*, *Clarias batrachus*, *Labeo rohita*, *Oreochromis mossambicus*, *Tor putitora*, *Cyprinus carpio*, and *Catla catla* etc. [31-38]. Pesticides exposure also alters the swimming speed and growth of the fish.

1.2.2. b. Impacts on fish reproduction

So far, scientific literature has evidenced toxicological influence in the form of reproductive abnormalities among various living beings. For example, pesticide exposure led to reproductive dysfunction in *Salmo salar*, *Salmo trutta* and *Mystus vitatus* [39-40], and reduced fertility in *Pseudorasbora*. The Physiological disturbance is also observed in some fish species [41]. Different studies also revealed the developmental toxicological effects of pesticides in fish [42-43]. Pesticides including dichlorvos, diazinon, carbufuran, endosulfan, and malathion, atrazine, and deltamethrin led to pathological toxicity in different organs of the fish including gills, kidney, liver, and muscles.

The pathological effects were necrosis, orientation loss of plates, cytoplasm granularity, cells shrinkage, oocytes increase, damage to collecting duct, follicular cells degeneration, nuclear materials shrinkage, vacuolation, change in the size of tubular line, nuclear pycnotic alterations, glomerulus degeneration, and rupturing of epithelial lining in *Glossogobius giuris*, *Gambusia affinis*, *L. rohita*, *Corydoras paleatus*, *Lepomis macrochirus*, *C. carpio communis*, *C. punctatus*, *O. niloticus*, *C. mrigala*, *Aphanius dispar*, *Glossogobius giuris*, *Macrobrachium malcolmsonii*, *O. mossambicus* and *Heteropneustes fossilis* [44-54].

1.2.2. C. Impacts on fish hematology

Hematological effects including anemias, neutropenia, lymphocytopenia, decreased R.B.Cs. count, etc., of blood parameters related to pesticide toxicity were observed in *T. putitora*, *O. niloticus*, *O. mossambicus*, *C. punctatus*, *C. batrachus*, *C. carpio*, *Puntius ticto* and *Onchorhynchus mykiss* [55 – 66].

1.2.2. d. Impacts on fish enzymes

Exposure to pesticides led to changes in the activities of different enzymes such as citrate synthase (CS), glucose 6-phosphate phosphate dehydrogenase (G6-PDH), lactate dehydrogenase (LDH), L-Keto acid glutaminase and in the brain, gills, liver and muscles of *C. batrachus* and *L. rohita* [67-70]. Pesticide exposure led to AChE (acetylcholine esterase) inhibition in different fish species including *O. massambicus*, *L. rohita*, *C. carpio*, and *Leporinus obtusidens*, *Jenynsia multidentata* and *Odontesthes hatcheri* etc., leading to neurotoxicity [71-73]. Pesticide exposure altered the feeding biology of *Puntius stigma*, and *Oryzias carnaticus* [74-75], as well as led to endocrine disruption in *O. mykiss* and *L. rohita* [76- 77].

Pesticides altered the activities of different enzymes of the antioxidant enzyme system including glutathione-s-transferase, glutathione reductase, superoxide dismutase, lipid peroxidase, peroxidase and catalase in *T. Putitora*, *C. gariepinus*, *L. macrochirus*, *Hoplias malabaricus*, *O. niloticus*, *L. rohita*, *O. hatcheri* and *J. multidentata* [78-84].

Pesticides exposure led to immunotoxicity in some fish species by reducing the phagocytosis, the number of granulocytes, lymphocytes, and leukocytes, inhibiting proliferation of B, T and screening cells (antibody) and ultimately decreasing the resistance of fish to infections and diseases [85-90]. Exposure to pesticides led to changes in proximate composition of *C. carpio*, *H. fossilis*, *C. gariepinus*, *Colisafasciatus*, *C. batrachus*, *O. niloticus*, *Puntiusticto*, and *L. rohita* via increase in cholesterol and ascorbic acid or decrease in protein, albumin and glycogen [91-99].

1.2.2. e. Genotoxic impact of pesticides in fish

Pesticides exposure led to genotoxicity in *L. rohita*, *C. mrigala* and *C. auratus* [100-101]. Pesticide exposure induced carcinogenic, mutagenic and genotoxic effects in fish and altered the genetic material like mediation of centromeric gaps, attenuations, stubbed arms of chromosomes, extra fragments, pycnosis, chromatid breaks and gaps, etc., consequent alteration of DNA replication, ultimately leading to cell proliferation and mutations [102].

2. Heavy metals

Heavy metal pollution sources of Egyptian water are mainly the agriculture drainage, sewage disposal, pesticides in water and industrial effluents and fertilizers [103-104]. Heavy metals are frequently released from human-made activities and natural sources into the aquatic ecosystems and accumulated in fish which situated at the top of the food chain and can accumulate large amounts of heavy metals [105].

To our knowledge, heavy metals delivered many unfavorable health effects. However, the vulnerability to heavy metal is common and persistent and even increasing in some parts of the world, mostly in rural districts. Metals are divided into two major classes: the first one is the essentials that function a proper role inside the body, while others like lead, cadmium and arsenic are not essentials and may be toxic [106].

2.1. Heavy metals and water pollution

Industrial effluents and metal industry constitute a real threat to the aquatic ecosystems. It represents about 50% of the total waste discharges and [107]. Serious concerns were found due to the presence of heavy metals in waterproducing serious influences on plant and animal life. Lead, cadmium and many other elements cause extreme toxicity at even trace levels [108]. Recent studies have shown that the natural habitat of fish and other marine organism which is exposed to human activities are under ecological pressure [109].

Taken into consideration, fish are the most susceptible organisms in the aquatic ecosystem to toxic substances that is accumulated in water [110]. Fish usually accumulate heavy metals in their bodies either directly through daily water consumption or through the gills, skin, and digestive tract [111]. Eventually, human health is exposed to risk through fish intake as fish is a main ingredient in human food [112]. Therefore, heavy metals accumulation in water and eventually fish has been a great worldwide problem; not only due to bad fish health concerns, but also due to the human health risks associated with fish consumption [113].

Water pollution by heavy metals creates negative impacts as follows:

2.1.1. Residues of heavy metals in water

In the aquatic ecosystem, heavy metals are the most important form of pollution due to their bioaccumulation by marine organisms [114]. The metabolic processes inside the living body usually need some trace elements, which are assimilated by marine organisms. Although, these living organisms form complexes with organic substances that can result in concentrations up to 1000 times higher than their assimilation and fixation in tissues and so, these metals become toxic to organisms [115].

In the aquatic ecosystem, water, suspended solids, sediments and biota are the main environmental components where trace elements are divided [116]. Sediment contamination is considered the major environmental problems in ecosystems. The pollution status of the environment is properly

assessed through sediment analysis [117]. Generally, the variations in metals concentration in water are depending on several factors as climate, soil type, pH, redox potential and dilution capacity [118].

A previous study analyzed the most environmental metals including; Pb, Cd, Ni, Co, Cu, and Zn in twenty five samples which were collected from both water and surficial bottom sediments of El-Manzala Lake. The study displayed that Cd caused a serious pollution due to the massive use of phosphate fertilizers. The mean Cd content in the area of the study was 17.5 ppm, which is about 36 fold the Maximum Permissible Limit (MPL=0.5 ppm) of soil. The major concentration of Cd is 22.3 ppm, which is more than 45 fold the MPL. Cadmium is more mobile in aquatic environments than most other heavy metals. The areas around Port Said and El-Serw drain show marked pollution by most of the studied heavy metals. The main reason for such pollution is the industrial activities and agricultural drains [119].

The main origin of Ni pollution in water was evaluated. Domestic wastewater effluents were found to be the major source, where, the drinking water and acidic beverages may dissolve Ni from pipes. Also, the advancement in industrialization progress increases the Ni discharge into the environment, where many nickel compounds are introduced in the commercial and industrial uses. It was found that water samples was greater than the authorized concentration for drinking water ($20 \mu\text{g L}^{-1}$) in the studied human-made activities impact points and reached $33.1 \mu\text{g L}^{-1}$ at the River Nile [120].

Iron concentration in fresh water was estimated [121]. The study concluded that iron concentration in water is managed by flow and redox conditions, the type and amount of dissolved organic matter and water pH. The results of the study deduced that Fe concentrations overstepped the aquatic life guidelines [122] where the anthropogenic activities in the Nile River have led to high concentrations of iron into its streams.

The water quality in El-Max fish farm was evaluated [123]. The results depicted the strong association between Zn and Cd in fish

farm water. The highest concentration of heavy metals Cd and Zn exerted adverse biological effects.

2.1.2. Impacts of heavy metals on marine life

Heavy metals can affect negatively on the growth rates, physiological functions, mortality and reproduction in fish [124].

2.1.3. a. Impacts on fish growth

Growth and maturation of fish larvae and juveniles is very fast. In the water polluted with heavy metals, fish growth may be inhibited. Inhibition of growth is one of the most distinct symptoms of metal toxicity on fish larvae. Therefore, environmental pollution is indicated by fish body length and mass [125].

2.1.2. b. Impacts on fish behavior

The data of many authors indicated that heavy metals reduce survival and growth of fish larvae [126-127]. Also, behavioral abnormalities such as impaired locomotors attitude resulting in increased susceptibility to predators were evident [128]. Also, fish behaviors may be changed by some heavy metals like Zinc (Zn). The main identified changes due to Zn include balance disorders, confused swimming, air guzzling, periods of dormancy and death [126].

Behavioral abnormalities like loss of equilibrium, jerky body movements, rolling, mucous secretion over the body, rapid opercular movements, difficulty in breathing and lethargic hyperactivity, erratic swimming, swallowing of air, rotations, convulsions and accelerated ventilation with rapid arrhythmic mouth movements after the exposure to several elements (Cd, Pb, Hg, Cu) in various fish species as *Tilapia mossambica*, *Eutroplus maculatus*, *Salvalinus fontinalis*, *Salmogairdneri*, and *Clarias batrachus* [127-131].

2.1.2. c. Impacts on fish reproduction

It is well established that many chemicals, both natural and man-made, may adversely affect wildlife, including decrease in fertility, disrupted hormone secretion or reproductive histopathology. Damaged ovarian follicles were observed in female mussels exposed to tetrabromodiphenyl ether [132].

A reduced gonad size with delayed sexual maturity in perch exposed to leachate from Swedish refuse dumps. Also, ovarian growth inhibition, reduction of egg weight and increased atresia, were depicted in white perch in areas polluted with domestic and industrial effluents. The discrimination of gonads and development of salmon were affected by sewage effluents [133]. Fish living in Bizerta lagoon, downstream of anthropogenic pollution have been found to exhibit an array of altered features in their reproductive development, including sperm abnormality and ovarian atresia [134].

The impact of diluted levels of polluted seawater from the Egyptian Mediterranean coast on reproductive, toxicological and hematological characteristics of *Siganus rivulatus* was investigated. Contaminated water has a harmful effect on gonads differentiation, changed endocrine haemostasis, and testosterone and progesterone levels reduction in females. While in male, progesterone level increased. Necrosis of spermatogenic cells and atresia of developing oocytes are pronounced at levels of 10 and 15 ml L⁻¹ polluted seawater [135].

2.1.2. d. Impacts on fish hemopoietic system

The damage and changes of the hemopoietic system of fish is mainly due to water-borne pollutants [136]. Several previous studies [137-139] investigated the sub-lethal effect of some heavy metals (Pb, Cd, and Cu) on *Siganusrivulatus*. The results were concluded in some changes in the morphology of the R.B.Cs such as tear drop-like cells, acanthocytes, and sickle cells. [140] found that R.B.Cs of *Clariaslazera* poisoned with lead were deteriorated, as clumping of chromatin material and disintegration of their cellular and nuclear membrane. It was examined that the exposure of Indian carp *Caltacalta* to copper [141]. The results were concluding in shrinkage of the configuration of the red blood cells with slight anisocytosis and an inclination to overlap. It was found that upon the exposure of *Tilapia niloticato* sub-lethal concentration of copper resulted in significant increase in erythrocyte count and haemoglobin content was observed [142].

2.1.2. e. Impacts on fish musculature

The tissue of greatest concern regarding heavy metal accumulation, from human standpoint, is the musculature. Most of the heavy metals bio-accumulates poorly in fish musculature only in the heavily contaminated environment [143].

the influence of the heavy metal pollution in the Lake Mariuot on some musculature biochemical constituents of *Oreochromis niloticus* fish was studied [143]. The study results showed a significant elevation in the lead, cadmium, copper, zinc and iron levels in musculature comparing with reference fish collected from a relatively clean aquaculture. The fish musculature protein, lipid, calcium, phosphorus and the amino acids, methionine, lysine and cystine concentrations were significantly decreased. The accumulation of heavy metals in *Oreochromisniloticus* fish followed the order: Mn>Zn>Pb>Cu. The highest levels of the heavy metals were found in the intestine and the lowest was found in the muscles [144]. Cu concentrations in the muscle of fish were below the maximum permissible limit, however, Mn, Pb and Zn exceeded the permissible limits. Total proteins, total lipids and activities of ALT and AST were significantly lower in the muscles of the studied fish from the River Nile in Egypt.

2.1.2. f. Impacts on immune system of fish

Phagocyte activity of fish macrophages has been inhibited by Lead, mercury and cadmium and so the cell mediated immune response has been an inhibited. The humoral immune response also affected due to these metals which is manifested by low levels of antibodies and high mortality rates in fish exposed to these metals than in the control fish after experimental infection by *Pseudomonas flourscens*. Immune response by these metals provides opportunities for the entry of pathogens and developing of many diseases in fish [145]. Heavy metals take great attention and special importance due to their increased toxic effects on fish as they affect live, growth, development and reproduction [145-146].

There was significant ($P < 0.01$) difference in WBCs of fish collected from the different studied sites along Borollus Lake. Moreover,

fish collected from the western and eastern sites of the lake had a significant increase in WBCs count, than that of fish collected from El-Boughaz opening. The reported leukocytosis in this study may be due to elevated leukocyte mobilization to conserve the body against infections in metals-affected tissue [147]. The elevation in the number of WBCs of fish was suggested to be due to alteration in defense mechanism to manage the highly toxic and the bioaccumulated heavy metals in fish organs as reported by [148-151].

2.1.2. f. Impacts on fish antioxidant enzymes

Heavy metals are recognized to decrease the antioxidant enzymes activities of (e.g. superoxide dismutase, Catalase) [152]. Heavy metal give rise to oxidative stress that is manifested by redox cycling and interaction with organic pollutants contribute to from aquatic pollution. Oxidative stress has been identified as a causative agent in a number of pathologies in fish resulting from reactive oxygen species (ROS) [153]. Oxidative stress occurs as a conclusion of excessive reactive oxygen species (ROS) and nitrogen species (RNS) release and it is improved by endogenous antioxidant enzyme activity and exogenous dietary antioxidants [154]. Cells that manage oxidative stress show many alterations due to the effect of ROS on serious structures in the body (lipids, proteins and DNA) [155]. Oxidative stress resulted from pollutants is usually manifested by elevated levels of lipid peroxides (LPO) (products of oxidative damage) and Malonaldehyde (MDA) and subsequent increase in defense enzymes (GSH, SOD activity and CAT ase) in response to the stress [156] or decrease due to overwhelming effect of the pollutants [157]. Nitric oxide is a highly unique and ubiquitous signaling molecule which is known to play variable physiological functions including those of adaptation to various stresses. Metal bioaccumulation is found to be in the same line with increased LPO and MDA levels and certain biomarkers of oxidative stress as surrogate bio-indicators of aquatic pollution in *Clarias gariepinus* [158].

Conclusion

This review highlights the causes and consequences of the water sources

contamination on fish organs in Egypt. There was a strong evidence of a correlation between water contaminants in different fish tissue and those of the surface water of the polluted areas. All the aforementioned sources of pollution affect the physicochemical characteristics of the water, sediments and biological components, thus negatively affecting the quality and quantity of fish stocks.

Conflict of interest

The authors have no conflict of interest to declare.

References

- [1]Deshpande, S. and Aher, K. (2012): Evaluation of groundwater quality and its suitability for drinking and agriculture use in parts of Vaijapur, District Aurangabad, MS, India. Res. J. Chem. Sci., 2(1): 25-31.
- [2]UNESCO, W. (2003): Water for people water for life: UN World Water Development Report. Paris: UN.
- [3]Dixit, R.C.; Verma, S.R.; Nitnaware, V. and Thacker, N.P. (2003): Heavy metals contamination in surface and groundwater supply of an Urban city. Indian J. Environ. Health, 45 (2): 107-112.
- [4]Cook, J.L.; Baumann, P.; Jackman, J.A. and Stevenson, D. (1995): Pesticides characteristics that affect water quality. College Station, TX: Texas Agricultural Extension Service, Texas A & M University. Available: http://insects.tamu.edu/extension/bulletins/water/water_01.html [accessed 23 October 2009].
- [5]Gunkel, G.; Kosmol, J.; Sobral, M.; Rohn, H.; Montenegro, S. and Aureliano, J. (2007): Sugar cane industry as a source of water pollution - case study on the situation in Ipojuca River, Pernambuco, Brazil. Water Air Soil Pollut, 180 (1-4): 261-269.
- [6]Ahmad, I.; Hassan, S. and Ahmad, I. (2013): Bacteriological quality analysis of drinking water of rural areas of Peshawar, Pakistan. American-Eurasian J. Agric. & Environ. Sci., 13 (9): 1202-1206.

- [7] Ullah, R.; Zuberi, A.; Tariq, M. and Ullah, S. (2014): Acute toxic effects of cypermethrin on hematology and morphology of liver, brain and gills of Mahseer (*Tor putitora*). *Int. J. Agri. Biol.*, 17(1), 199-204.
- [8] Dural, M., Göksu, L.Z.M.; Özak, A. A. and Derici, B. (2006): Bioaccumulation of some heavy metals in different tissues of *Dicentrarchus labrax* L, 1758, *Sparus aurata* L, 1758 and *Mugil cephalus*, L, 1758 from the Camlik Lagoon of the eastern coast of Mediterranean (Turkey). *Environ. Monit. Assess.* 18 (1-3):65-74.
- [9] Erdoğrul, Z. and Ates, D.A. (2006): Determination of cadmium and copper in fish samples from Sir and Menzelet dam lake Kahramanmaraş, Turkey. *Environ. Monit. Assess.* 117 (1-3):281-290.
- [10] Liu, W.-X.; Wang, Y.; He, W.; Qin, N.; Kong, X.-Z.; He, Q.-S., Yang, B.; Yang, C.; Jiang, Y.-J. and Jorgensen, S.E. (2016): Aquatic biota as potential biological indicators of the contamination, bioaccumulation and health risks caused by organochlorine pesticides in a large, shallow Chinese lake (Lake Chaohu). *Ecol. Indic.*, 60: 335-345.
- [11] Marigoudar, S.R.; Nazeer Ahmed, R. and David, M. (2012): Impact of Cypermethrin on Behavioural Responses in the Freshwater Teleost, *Labeo rohita* (Hamilton).. *World Journal of Zoology* (WJZ) 4 (1): 19-23.
- [12] Fevery, D.; Houbraken, M. and Spanoghe, P. (2016): Pressure of non-professional use of pesticides on operators, aquatic organisms and bees in Belgium. *Sci. Total Environ.*, 550: 514-521.
- [13] Sarkar, S.; Bhattacharya, B.; Bhattacharya, A.; Chatterjee, M.; Alam, A.; Satpathy, K. and Jonathan, M. (2008): Occurrence, distribution and possible sources of organochlorine pesticide residues in tropical coastal environment of India: an overview. *Environ. Int.*, 34(7), 1062-1071.
- [14] Shanbehzadeh, S.; Vahid Dastjerdi, M.; Hassanzadeh, A. and Kiyanzadeh, T. (2014): Heavy metals in water and sediment: a case study of Tembi River. *J. Environ. Public Health*, 2014: Article ID 858720, 5 pages..
- [15] Hua, J. and Relyea, R. (2014): Chemical cocktails in aquatic systems: Pesticide effects on the response and recovery of > 20 animal taxa. *Environ. Pollut.*, 189, 18-26.
- [16] Kasmin, S. (2010): Enforcing ship-based marine pollution for cleaner sea in the Strait of Malacca. *Environment Asia*, 3: 61-65.
- [17] Cimino, A.M.; Boyles, A.L.; Thayer, K.A. and Perry, M.J. (2017): Effects of neonicotinoid pesticide exposure on human health: a systematic review. *Environ. Health Perspect.*, 125(2): 155-162.
- [18] Levi, S.; Hybel, A.M.; Bjerg, P.L. and Albrechtsen, H.J. (2014): Stimulation of aerobic degradation of bentazone, mecoprop and dichlorprop by oxygen addition to aquifer sediment. *Sci. Total Environ.*, 473: 667-675.
- [19] Schäfer, R.B.; Gerner, N.; Kefford, B.J.; Rasmussen, J.J.; Beketov, M.A.; de Zwart, D.; Liess, M. and von der Ohe, P.C. (2013): How to characterize chemical exposure to predict ecologic effects on aquatic communities? *Environ. Sci. Technol.*, 47(14): 7996-8004.
- [20] Shuman-Goodier, M.E. and Propper, C.R. (2016): A meta-analysis synthesizing the effects of pesticides on swim speed and activity of aquatic vertebrates. *Sci. Total Environ.*, 565, 758-766.
- [21] Barakat, A.O.; Kim, M.; Qian, Y. and Wade, T.L. (2002): Organochlorine pesticides and PCB residues in sediments of Alexandria Harbour, Egypt. *Mar. Pollut. Bull.*, 44(12): 1426-1434.
- [22] Barakat, A.O.; Khairy, M. and Aukaily, I. (2013): Persistent organochlorine pesticide and PCB residues in surface sediments of Lake Qarun, a protected

- area of Egypt. *Chemosphere*, 90(9): 2467-2476.
- [23] El Nemr, A. and El-Sadaawy, M.M. (2016): Polychlorinated biphenyl and organochlorine pesticide residues in surface sediments from the Mediterranean Sea (Egypt). *Int. J. Sediment Res.*, 31(1): 44-52.
- [25] Ragab, S.; El Sikaily, A. and El Nemr, A. (2016): Concentrations and sources of pesticides and PCBs in surficial sediments of the red sea coast, Egypt. *Egypt J. Aquat. Res.*, 42(4): 365-374.
- [26] Maskaoui, K.; Zhou, J.; Zheng, T.; Hong, H. and Yu, Z. (2005): Organochlorine micropollutants in the Jiulong River estuary and western Xiamen Sea, China. *Mar. Pollut. Bull.*, 51(8-12): 950-959.
- [27] Helfrich, L.A.; Weigmann, D.L.; Hipkins, P.A. and Stinson, E.R. (1996): Pesticides and aquatic animals: a guide to reducing impacts on aquatic systems. VCE Publications / 420 / 420-013. <http://pubs.ext.vt.edu/420/420-013/420-013.html>
- [28] Gill, R.J. and Raine, N.E. (2014): Chronic impairment of bumblebee natural foraging behaviour induced by sublethal pesticide exposure. *Funct. Ecol.*, 28(6): 1459-1471.
- [29] Pérez, J.; Domingues, I.; Monteiro, M.; Amadeu, M.; Soares, M. and Loureiro, S. (2013): Synergistic effects caused by atrazine and terbuthylazine on chlorpyrifos toxicity to early-life stages of the zebrafish *Danio rerio*. *Environ. Sci. Pollut. Res. Int.*, 20(7), 4671-4680.
- [30] Rasmussen, J.J.; Nørum, U.; Jerris, M.R.; Wiberg-Larsen, P.; Kristensen, E.A. and Friberg, N. (2013): Pesticide impacts on predator-prey interactions across two levels of organisation. *Aquat. Toxicol.*, 140: 340-345.
- [31] Murthy, K.S.; Kiran, B. and Venkateshwarlu, M. (2013): A review on toxicity of pesticides in Fish. *International Journal Open Science Research (IJSR)*, 1(1): 15-36.
- [32] Ilavazhahan, M.; Selvi, R.T. and Jayaraj, S. (2010): Determination of LC of the Bacterial Pathogen, Pesticide and. *Global Journal of Environmental Research (GJER)*, 4(2): 76-82.
- [33] Krian, A. and Jha, A. (2009): Acute toxicity and behavioural responses of herbicide (Herboclin) to the fish *Clarias batrachus* (Linn). *Indian J. Environ. Ecolplan*, 16(1): 185-188.
- [34] Marigoudar, S.R.; Ahmed, R.N. and David, M. (2009): Cypermethrin induced respiratory and behavioural responses of the freshwater teleost, *Labeo rohita* (Hamilton). *Vet. Arhiv*, 79(6): 583-590.
- [35] Nagaraju, B.; Sudhakar, P.; Anitha, A.; Haribabu, G. and Rathnamma, V. (2011): Toxicity evaluation and behavioural studies of fresh water fish *Labeo rohita* exposed to Rimon. *Int. J. Res. Phar. Biomed. Sci.*, 2: 2229-3701..
- [36] Nwani, C.D.; Lakra, W.S.; Nagpure, N.S.; Kumar, R.; Kushwaha, B. and Srivastava, S.K. (2010): Toxicity of the herbicide atrazine: effects on lipid peroxidation and activities of antioxidant enzymes in the freshwater fish *Channa punctatus* (Bloch). *Int. J. Environ. Res. Public Health.*, 7(8), 3298-3312.
- [37] Rani, S. and Venkataramana, G. (2012): Effects of the organophosphorous Malathion on the branchial gills of a freshwater fish *Glossogobius giuriss* (Ham). *Int. J. Sci. Nat* 3(2): 324-330.
- [38] Scott, G.R. and Sloman, K.A. (2004): The effects of environmental pollutants on complex fish behaviour: integrating behavioural and physiological indicators of toxicity. *Aquat. Toxicol.*, 68(4), 369-392.
- [39] Ullah, R.; Zuberi, A.; Ullah, S.; Ullah, I. and Dawar, F.U. (2014): Cypermethrin induced behavioral and biochemical changes in mahseer, *Tor putitora*. *J Toxicol Sci.*, 39(6), 829-836.
- [40] Jaensson, A.; Scott, A.P.; Moore, A.; Kylin, H. and Olsén, K.H. (2007): Effects of a pyrethroid pesticide on endocrine responses to female odours

- and reproductive behaviour in male parr of brown trout (*Salmo trutta* L.). Aquat. Toxicol., 81(1), 1-9.
- [41] Moore, A. and Waring, C.P. (2001): The effects of a synthetic pyrethroid pesticide on some aspects of reproduction in Atlantic salmon (*Salmo salar* L.). Aquat. Toxicol., 52(1): 1-12.
- [42] Chandra, G.; Bhattacharjee, I.; Chatterjee, S. and Ghosh, A. (2008): Mosquito control by larvivorous fish. Indian J Med Res., 127(1): 13-28.
- [43] Corvi, M.M.; Stanley, K.A.; Peterson, T.S.; Kent, M.L.; Feist, S.W.; La Du, J.K.; Volz, D.C.; Hosmer, A.J. and Tanguay, R.L. (2012): Investigating the impact of chronic atrazine exposure on sexual development in zebrafish. Birth Defects Res. B Dev. Reprod. Toxicol., 95(4): 276-288.
- [44] Xu, C.; Tu, W.; Lou, C.; Hong, Y. and Zhao, M. (2010): Enantioselective separation and zebrafish embryo toxicity of insecticide beta-cypermethrin. J. Environ. Sci., 22(5), 738-743.
- [45] Ba-Omar, T.; Al-Jardani, S. and Victor, R. (2011): Effects of pesticide temephos on the gills of *Aphanius dispar* (Pisces: Cyprinodontidae). Tissue Cell, 43(1): 29-38.
- [46]. Benli, A.Ç.K. and Özkul, A. (2010): Acute toxicity and histopathological effects of sublethal fenitrothion on Nile tilapia, *Oreochromis niloticus*. Pest. Biochem. Physiol., 97(1): 32-35.
- [47] David, M.; Mushigeri, S.; Shivakumar, R. and Philip, G. (2004): Response of *Cyprinus carpio* (Linn) to sublethal concentration of cypermethrin: alterations in protein metabolic profiles. Chemosphere, 56(4): 347-352.
- [48] Deka, S. and Mahanta, R. (2012): A study on the effect of organophosphorus pesticide malathion on hepato-renal and reproductive organs of *Heteropneustes fossilis* (Bloch). The Science Probe, 1(1): 1-13.
- [49] Dutta, H. and Meijer, H. (2003): Sublethal effects of diazinon on the structure of the testis of bluegill, *Lepomis macrochirus*: a microscopic analysis. Environ Pollut, 125(3), 355-360.
- [50] Fanta, E.; Rios, F.S.A.; Romão, S.; Vianna, A.C.C. and Freiberger, S. (2003): Histopathology of the fish *Corydoras paleatus* contaminated with sublethal levels of organophosphorus in water and food. Ecotoxicol. Environ. Saf., 54(2): 119-130.
- [51] Johal, M.S.; Sharma, M.L. and Ravneet (2007): Impact of low dose of organophosphate, monocrotophos on the epithelial cells of gills of *Cyprinus carpio communis* Linn.-SEM study. J Environ Biol, 28(3): 663-667.
- [52] Aniladevi Kunjamma, K.P.; Philip, B.; Bhanu, S.V. and Jose, J. (2008): Histopathological effects on *Oreochromis mossambicus* (Tilapia) exposed to chlorpyrifos. J Environ Res Dev, 2(4): 553-559.
- [53] Manosathiyadevan, M.; Selvisabhanayakam, V. and Divya, M. (2012): Morphological alterations and biochemical contents of the testis of adult male freshwater prawn *Macrobrachium malcolmsonii*. Ind J Fund Appl Lif Sci, 2(4): 104-113.
- [54] Mir, F.A.; Shah, G.M.; Jan, U. and Mir, J.I. (2012): Studies on influences of sublethal concentrations of organophosphate pesticide dichlorvos (DDVP) on Gonadosomatic Index (GSI) of female common carp, *Cyprinus carpio communis*. Am-Euras J Toxicol Sci, 4(2): 67-71.
- [55] Velmurugan, B.; Selvanayagam, M.; Cengiz, E.I. and Unlu, E. (2009): Histopathological changes in the gill and liver tissues of freshwater fish, *Cirrhinus mrigala* exposed to dichlorvos. Braz Arch Biol echnol, 52(5): 1291-1296.
- [56] Agrahari, S.; Pandey, K.C. and Gopal, K. (2007): Biochemical alteration induced by monocrotophos in the blood plasma

- of fish, *Channa punctatus* (Bloch). Pest Biochem Physiol, 88(3): 268-272.
- [57] Banaee, M. (2013): Physiological dysfunction in fish after insecticides exposure. In Trdan S. editor. Insecticides-Development of Safer and More Effective Technologies. Croatia: InTech.
- [58] Banaee, M.; Mirvagefei, A.; Rafei, G. and Majazi Amiri, B. (2008): Effect of sub-lethal diazinon concentrations on blood plasma biochemistry. Int J Environ Res, 2(2): 189-198.
- [59] Banaee, M.; Sureda, A.; Mirvaghefi, A. and Ahmadi, K. (2011): Effects of diazinon on biochemical parameters of blood in rainbow trout (*Oncorhynchus mykiss*). Pest biochem physiol, 99(1): 1-6.
- [60] Devi, P.; Baruah, D.; Baruah, B. and Borkotoki, A. (2008): Impact of endosulfan on some haematological parameters of *Channa punctatus* (Bloch). Pollut Res, 27(3): 485-488.
- [61] Far, M.S.; Roodsari, H.V.; Zamini, A.; Mirrasooli, E. and Kazemi, R. (2012): The effects of diazinon on behavior and some hematological parameters of fry rainbow trout (*Oncorhynchus mykiss*). World J Fish Marine Sci, 4(4): 369-375.
- [62] Gautam, R.K. and Kumar S. (2008): Alteration in haematology of *Channa punctatus* (Bloch). J Exp Zool, India, 11(2): 309-310.
- [63] Kavitha, P. and Rao, J.V. (2009): Sub-lethal effects of profenofos on tissue-specific antioxidative responses in a Euryhaline fish, *Oreochromis mossambicus*. Ecotoxicol environ saf, 72(6): 1727-1733.
- [64] Matos, P.; Fontai, A.; Peixoto, F.; Carrola, J. and Rocha, E. (2007): Biochemical and histological hepatic changes of Nile tilapia *Oreochromis niloticus* exposed to carbaryl. Pest Biochem and Physiol, 89(1): 73-80.
- [65] Satyanarayan, S.; Bejankiwar, R.S.; Chaudhari, P.R.; Kotangale, J.P. and Satyanarayan, A. (2004): Impact of some chlorinated pesticides on the haematology of the fish *Cyprinus carpio* and *Puntius ticto*. J Environ Sci, 16(4): 631-634.
- [66] Velisek, J.; Svobodova, Z. and Machova, J. (2009): Effects of bifenthrin on some haematological, biochemical and histopathological parameters of common carp (*Cyprinus carpio* L.). Fish physiol biochem, 35(4): 583-590.
- [67] Venkateswara Rao, J.; Kavitha, P.; Jakka, N.M.; Sridhar, V. and Usman, P.K. (2007): Toxicity of organophosphates on morphology and locomotor behavior in brine shrimp, *Artemia salina*. Arch environ contamin toxicol, 53(2): 227-232.
- [68] Mastan, S. and Shaffi, S. (2010): Sub-lethal Effect of Pesticides on the Distribution of Glutaminases in the Brain of *Labeo rohita* (Ham.). Int J Toxicol, 7(2): 1-6.
- [69] Thenmozhi, C.; Vignesh, V.; Thirumurugan, R. and Arun, S. (2011): Impacts of malathion on mortality and biochemical changes of freshwater fish *Labeo rohita*. Iran. J Environ Health Sc Eng, 8(4): 325-332.
- [70] Tripathi, G. and Verma, P. (2004): Endosulfan-mediated biochemical changes in the freshwater fish *Clarias batrachus*. Biomed Environ Sci, 17(1): 47-56.
- [71] Ullah, S.; Hasan, Z. and Dhama, K. (2016): Toxic effects of endosulfan on behaviour, protein contents and antioxidant enzyme system in gills, brain, liver and muscle tissues of rohu, *Labeo rohita*. Int J pharmacol, 12(1): 1-10.
- [72] Bibi, N.; Zuberi, A.; Naeem, M.; Ullah, I.; Sarwar, H. and Atika, B. (2014): Evaluation of acute toxicity of karate and its sub-lethal effects on protein and acetylcholinestrace activity in *Cyprinus carpio*. Int J Agricul Biol, 16(4): 731-737.
- [73] Guerreño, M.; López Armengol, M.F.; Luquet, C.M. and Venturino, A. (2016):

- Comparative study of toxicity and biochemical responses induced by sublethal levels of the pesticide azinphosmethyl in two fish species from North-Patagonia, Argentina. *Aquat Toxicol*, 177: 365-372.
- [74] Guo, J.X.; Wu, J.J.; Wright, J.B. and Lushington, G.H. (2006): Mechanistic insight into acetylcholinesterase inhibition and acute toxicity of organophosphorus compounds: a molecular modeling study. *Chem res toxicol*, 19(2): 209-216.
- [75] Marigoudar, S.R.; Ahmed, R.N. and David, M. (2009): Cypermethrin induced: in vivo inhibition of the acetylcholinesterase activity in functionally different tissues of the freshwater teleost, *Labeo rohita* (Hamilton). *Toxicol Environ Chem*, 91(6): 1175-1182.
- [76] Bhandare, R.; Pathan, T.; Shinde, S.; More, P. and Sonawane, D. (2011): Toxicity and behavioural changes in fresh water fish *Puntius stigma* exposed to pesticide (Rogor). *Am-Euras. J Toxicol Sci*, 3(3): 149-152.
- [77] Ravindran, K.J.; Daniel, R.; Kumari, S.; George, S. and Eapen, A. (2012): Effect of agricultural pesticides, Hostathion and Kitazin on the larvivorsity of the Carnatic rice fish, *Oryzias carnaticus* (Jerdon, 1849). *Amer Eur J Toxicol Sc*, 4(2): 56-59.
- [78] Dey, C. and Saha, S. K. (2014): A comparative study on the acute toxicity bioassay of dimethoate and lambda-cyhalothrin and effects on thyroid hormones of freshwater teleost fish *Labeo rohita* (Hamilton). *Int J Environ Res*, 8(4): 1085-1092.
- [79] Hoeger, B.; Hitzfeld, B.; Köllner, B.; Dietrich, D.R., and van den Heuvel, M.R. (2005): Sex and low-level sampling stress modify the impacts of sewage effluent on the rainbow trout (*Oncorhynchus mykiss*) immune system. *Aquat Toxicol*, 73(1): 79-90.
- [80] Elia, A.; Waller, W.T. and Norton, S.J. (2002): Biochemical responses of bluegill sunfish (*Lepomis macrochirus*, Rafinesque) to atrazine induced oxidative stress. *Bull Environ Contam Toxicol*, 68(6): 809-816.
- [81] Jiraungkoorskul, W.; Upatham, E.S.; Kruatrachue, M.; Sahaphong, S.; Vichasri-Grams, S. and Pokethitiyook, P. (2003): Biochemical and histopathological effects of glyphosate herbicide on Nile tilapia (*Oreochromis niloticus*). *Environ Toxicol*, 18(4): 260-267.
- [82] Milaeva, E.R. (2006): The role of radical reactions in organomercurials impact on lipid peroxidation. *J inorg biochem*, 100(5-6): 905-915.
- [83] Muthukumaravel, K.; Sivakumar, B.; Kumarasamy, P. and Govindarajan, M. (2013): Studies on the toxicity of pesticide monocrotophos on the biochemical constituents of the freshwater fish, *Labeo rohita*. *Int J Curr Biochem Biotechnol*, 2(10): 20-26.
- [84] Filipak Neto F.; Zanata, S.M.; Silva de Assis H.C.; Nakao, L.S.; Randi, M.A. and Oliveira Ribeiro C.A. (2008): Toxic effects of DDT and methyl mercury on the hepatocytes from *Hoplias malabaricus*. *Toxicol in Vitro*, 22(7): 1705-1713.
- [85] Nwani, C.D.; Nagpure, N.S.; Ravindra, K.; Basdeo, K.; Pavan, K. and Lakra, W.S. and (2010): Lethal concentration and toxicity stress of Carbosulfan, Glyphosate and Atrazine to freshwater air breathing fish *Channa punctatus* (Bloch). *Int Aquat Res*, 2(2): 105-111.
- [86] Shanker, G.; Syversen, T.; Aschner, J.L. and Aschner, M. (2005): Modulatory effect of glutathione status and antioxidants on methylmercury-induced free radical formation in primary cultures of cerebral astrocytes. *Brain Res Mol Brain Res*, 137(1-2): 11-22.
- [87] Girón-Pérez, M.I.; Santerre, A.; Gonzalez-Jaime, F.; Casas-Solis, J.; Hernández-Coronado, M.; Peregrina-Sandoval, J.; Takemura, A. and Zaitseva, G. (2007): Immunotoxicity and hepatic function evaluation in Nile tilapia (*Oreochromis*

- niloticus*) exposed to diazinon. Fish Shellfish Immunol, 23(4): 760-769.
- [88] Iwanowicz, L.R.; Blazer, V.S.; McCormick, S.D.; VanVeld, P.A. and Ottinger, C.A. (2009): Aroclor 1248 exposure leads to immunomodulation, decreased disease resistance and endocrine disruption in the brown bullhead, *Ameiurus nebulosus*. Aquat Toxicol, 93(1): 70-82.
- [89] Kreutz, L.C.; Barcellos, L.J.G.; Marteninghe, A.; Dos Santos, E.D. and Zanatta, R. (2010): Exposure to sublethal concentration of glyphosate or atrazine-based herbicides alters the phagocytic function and increases the susceptibility of silver catfish fingerlings (*Rhamdia quelen*) to *Aeromonas hydrophila* challenge. Fish shellfish immunol, 29(4): 694-697.
- [90] Narra, M.R. (2016): Single and cartel effect of pesticides on biochemical and haematological status of *Clarias batrachus*: A long-term monitoring. Chemosphere, 144: 966-974.
- [91] Shelley, L.K.; Balfry, S.K.; Ross, P.S. and Kennedy, C.J. (2009): Immunotoxicological effects of a sub-chronic exposure to selected current-use pesticides in rainbow trout (*Oncorhynchus mykiss*). Aquat toxicol, 92(2): 95-103.
- [92] Tellez-Bañuelos, M.C.; Santerre, A.; Casas-Solis, J. and Zaitseva, G. (2010): Endosulfan increases seric interleukin-2 like (IL-2L) factor and immunoglobulin M (IgM) of Nile tilapia (*Oreochromis niloticus*) challenged with *Aeromona hydrophila*. Fish shellfish immunol, 28(2): 401-405.
- [93] Aguigwo, J.N. (2002): The toxic effect of cymbush pesticide on growth and survival of African catfish, *Clarias gariepinus* (Burchell). J Aquat Sci, 17(2): 81-84.
- [94] Borah, S. (2005): Effect of petroleum oil on biochemical constituents and enzyme activity in kidney and liver tissues of freshwater teleost fish, *Heteropneustes fossilis* (Bloch). J Nat Environ Poll Technol, 4(2): 227-232.
- [95] Bose, S.; Nath, S. and Sahana, S. (2011): Toxic impact of thiamethoxam on the growth performance and liver protein concentration of a freshwater fish *Oreochromis niloticus* (Trewavas). Ind J Fund Appl Life Sci, 1(4): 274-280.
- [96] Ganeshwade, R.; Dama, L.; Deshmukh, D.; Ghanbahadur, A. and Sonawane, S. (2012): Toxicity of endosulfan on freshwater fish *Channa striatus*. Trends Fish Res, 1(1): 29-31.
- [97] Jenkins, F.; Smith, J.; Rajanna, B.; Shameem, U.; Umadevi, K.; Sandhya, V. and Madhavi, R. (2003): Effect of sub-lethal concentrations of endosulfan on hematological and serum biochemical parameters in the carp *Cyprinus carpio*. Bull environ contam toxicol, 70(5): 993-997.
- [98] Jha, B. and Verma, B. (2002): Effect of pesticidal mixture on protein content in the freshwater fish *Clarias batrachus*. J Ecotoxicol Environ Monit, 12(3): 177-180.
- [99] Lakshmanan, S.; Rajendran, C. and Sivasubramaniyan (2013): "Impact of Dichlorvos on tissue glycogen and protein content in freshwater fingerlings, *Oreochromis mossambicus* (Peters)". Int j Res Environ Sci Techno, 3(1): 19-25.
- [100] Rajput, V.; Singh, S.K.; Kirti A. and Abhishek (2012): Comparative toxicity of Butachlor, Imidacloprid and Sodium fluoride on protein profile of the walking cat fish *Clarias batrachus*. J Appl Pharmacol Sci, 2(6): 121-124
- [101] Ullah, S.; Hasan, Z.; Zorriehzahra, M. and Ahmad, S. (2017): Diagnosis of endosulfan induced DNA damage in rohu (*Labeo rohita*, Hamilton) using comet assay. Iran J Fish Sci, 16(1): 138-149.
- [102] Çavaş, T. and Könen, S. (2007): Detection of cytogenetic and DNA damage in peripheral erythrocytes of goldfish (*Carassius auratus*) exposed to a glyphosate formulation using the

- micronucleus test and the comet assay. *Mutagenesis* 22(4): 263-268.
- [103] Banaee, M.; Davoodi, M. and Zoheiri, F. (2013): Histopathological changes induced by paraquat on some tissues of gourami fish (*Trichogaster trichopterus*). *Open vet j*, 3(1): 36-42.
- [104] Santos, I. R.; Silva-Filho, E. V.; Schaefer, C. E.; Albuquerque-Filho, M. R. and Campos, L.S. (2005): Heavy metals contamination in coastal sediments and soils near the Brazilian Antarctic Station, King George Island. *Mar Pollut Bull.*, 50(2):185-194.
- [105] Singh, R.K.; Chavan, S.L.; and Sapkale, P.H. (2007): Heavy metal concentrations in water, sediments and body tissues of red worm (*Tubifex* spp.) collected from natural habitats in Mumbai, India. *Environ Monit Assess*, 129(1-3): 471-481.
- [106] Yilmaz, F.; Özdemir, N.; Demirak, A. and Tuna, A. L. [2007]: Heavy metal levels in two fish species *Leuscius cephalus* and *Lepomis gibbosus*. *Food Chem.*, 100: 830-835.
- [107] Jadhav, S.H.; Sarkar, S.N.; Patil, R.D. and Tripathi, H.C. [2007]: Effects of subchronic exposure via drinking water to a mixture of eight water-contaminating metals: a biochemical and histopathological study in male rats, *Arch. Environ. Contam. Toxicol.*, 53: [4]: 667-77.
- [108] El-Gohary, F.A. [1990]: Waste water management in Shubra El-Kheima. 2nd Egyptian seminar on industrial waste water management, Cairo, Egypt: 1-21.
- [109] Bertin, C. and Bourg, A.C.M. [1995]: Trends in the heavy metals content (Cd, Pb and Zn) of river sediments in the drainage basin of smelting activities. *Wat. Res.*, 29: 1729-1736.
- [110] Sen, I.; Shandi, A. and Shrivastava, V.S. [2011]: Study for determination of heavy metals in fish species of the River Yamuna (Delhi) by Inductively Coupled Plasma-Optical Emission Spectroscopy (ICP-OES). *Adv. Appl. Sci. Res.*, 2[2]: 161-166.
- [111] Alibabić, V.; Vahčić, N. and Bajramović, M. [2007]: Bioaccumulation of metals in fish of Salmonidae family and the impact on fish meat quality. *Environ. Monit. Assess.* 131: 349-364.
- [112] Burger, J.; Gaines, K. F.; Boring, C. S.; Stephens, W. L.; Snodgrass, J. and Dixon, C. McMahon, M.; Shukla, S.; Shukla, T. and Gochfeld, M. [2002]: Metal levels in fish from the Savannah River: potential hazards to fish and other receptors. *Environ. Res.*, 89: 85-97.
- [113] Begum, A.; Mustafa, A. I.; Amin, N.; Chowdhury, T. R.; Quraishi, S. B. and Banu, N. [2013]: Levels of heavy metals in tissues of shingi fish (*Heteropneustes fossilis*) from Buriganga River, Bangladesh. *Environ. Monit. Assess.* 185:5461-5469.
- [114] Ochieng, H.; Steveninck, E.S. and Wanda, F.M. [2008]: Mouthparts deformities in chironomides (dipteran) as indicators of heavy metal pollution in northern Lake Victoria, Uganda. *Afr. J. Aqua. Sci.*, 33[2]: 135-142.
- [115] Zeitoun, M.M. and Mehana, E. E. [2014]: Impact of Water Pollution with Heavy Metals on Fish Health: Overview and Updates. *Global Veterinaria*, 12 (2): 219-231, 2014.
- [116] Shakweer, L.M. and Abbas, M.M. [2005]: Effect of ecological and biological factors on the uptake and concentration of trace elements by aquatic organisms at Edku lake. *Egypt. J. Aquat. Res.* 31 [1]: 271-288.
- [117] Mucha, A.P.; Vasconcelos, M.T.S.D. and Bordalo, A.A. [2003]: Macrobenthic community in the Douro Estuary: relations with heavy metals and natural sediment characteristics. *Environ. Pollut.* 121: 169-180.
- [118] WHO, [1995]. Lead: Environmental Health Criteria. Vol. 165. Geneva:

- International Program on Chemical Safety (IPCS). World Health Organization, Switzerland
- [119] El-Badry, [2016]: Distribution of Heavy Metals in Contaminated Water and Bottom Deposits of Manzala Lake, Egypt, *J. Environ. Anal. Toxicol.*, 6[1]: 344-348.
- [120] Cempel, M. and Nickel, G. [2006]: Nickel: a review of its sources and environmental toxicology. *Polish J. Environ. Stud.*, 15 (3): 375-382.
- [121] Vuori, K. [1995]: Direct and indirect effects of iron on river ecosystems. *Ann. Zool. Fennici.*, 32: 317-329.
- [122] CCME (Canadian Council of Ministers of the Environment), [2007]: For the protection of aquatic life 2007. In: Canadian Environmental Quality Guidelines, 1999, Canadian Council of Ministers of the Environment, 1999, Winnipeg.
- [123] El-Zokm, G.M.; El-Gohary, S.E. and Abd El-Kalek, D.E. [2012]: Studies of some heavy metals in water and sediment in El-Max fish farm, Egypt. *World Appl. Sci. J.*, 18 [2]: 171-180.
- [124] Amundsen, P.A.; Staldivik, F.J.; Lukin, A.A.; Kashulin, N.A.; Popova, O.A. and Reshetnikov, Y.S. [1997]: Heavy metal contamination in freshwater fish from the border region between Norway and Russia. *Sci. Total. Environ.*, 201:211-224.
- [125] Sarnowski, P. and Jezierska B. [2007]: A new coefficient for evaluation of condition of fish: *Electron. J. Ichthyol.*, 2: 69-76.
- [126] Stomińska, I. and Jezierska, B. [2000]: The effect of Heavy metals on post embryonic development of COMMON CARP, *Cyprinus carpio L.*, *Archives of Polish Fisheries*, Vol. 8 Fasc., 1 119-128.
- [127] Kori, S.O and Ubogu O. E. [2008]. Sublethal hematological effects of zinc on the freshwater fish, *Heteroclaris sp.* (Osteichthyes:Clariidae), *Afr. J Biotech.*, 7[12]:2068-2073.
- [128] Ghatak, D. B. and Konar, S.K. [1990]: Acute toxicity of mixture of heavy metals cadmium, pesticides DDVP, detergent Parnol J and petroleum product n-heptane on fish , plankton and worm. *Environ. Ecol.*, 8[4]: 1239-1248.
- [129] Holecombe, G.W.; Benoit, D.A.; Leonard, E.N. and McKim, J. M. [1976]: Long term effects of lead exposure on three generations of brook trout, *Salvelinus fontinalis*. *J. Fish. Res. Board. Can.*, 33[8]: 1731-1741.
- [130] McLeod, J.C. and Pessah, E. [1973]: Temperature effects on mercury accumulation, toxicity and metabolic rate in rainbow trout, *Salmogairdneri*. *J. Fish. Res. Board. Can.*, 30[4]: 485-492.
- [131] Singh, H.S. and Reddy, T.V. [1990]: Effect of copper sulfate on haematology, blood chemistry and hepato somatic index of an Indian cat fish, *Heteropneustes fossilis* (Bloch) and its recovery. *Ecotoxicol. Environ. Saf.*, 20: 30-35.
- [132] Siddiqui, A.A.. and Arifa, N. [2011]: Toxicity of heavy metal copper and its effect on the behaviour of fresh water Indian cat fish, *Clarias batrachus* (Linn.). *Curr Biotica.*, 4[4]: 405-411.
- [133] Aarab, N.; Lemaire-Gony, S.; Unsuh, E. ; Hansen, P.D.; Andersen, O.K. and Narbonne, J.F. [2006]: Preliminary study of responses in mussel (*Mytilus edulis*) exposed to tetrabromodiphenyl ether. *Aquat. Toxicol.*, 78: S86-S92.
- [134] Noaksson, E.; Linderroth, M.; Tjarnlund, U. and Balk, L. [2005]: Toxicological effects and reproductive impairments in female perch (*Perca fluviatilis*) exposed to leachate from Swedish refuse dumps. *Aquat. Toxicol.*, 75: 162-177.
- [135] Afonso, L. O. B. ; Smith, J. L.; Ikonomou, M. G. and Devlin, R. H. [2002]: Y-chromosomal DNA markers for discrimination of chemical substances and effluent effects on sexual differentiation and gonadal

- development in salmon. Environ. Health Perspect., 110: 881-887.
- [136] Louiz, I.; Ben-Attia, M. and Ben-Hassine, O.K. [2009]: Gonadosomatic index and gonad histopathology of *Gobius niger* (Gobiidea, Teleost) from Bizerta lagoon (Tunisia): Evidence of reproduction disturbance. Fish. Res., 100: 266-273.
- [137] Moharram, S.G.; Wahbi, O.M. and El-Greisy, Z.A. [2011]: Effect of polluted water from the Egyptian Eastern Mediterranean coast on reproductive, toxicological and hematological characteristics of *Siganus rivulatus*. Pak J Biol Sci., 14(12): 668-681.
- [138] Gardner, G.R. and Yevich, P.P [1969b]: Toxicological effects of cadmium on *Fundulus heteroclitus* under various oxygen, pH., salinity and temperature regimes. Amer. Zool. g: 1096 (Abstr.).
- [139] Osman, M.A.; Ghaly, Y.S.; Afify, A.M.R.; El-Dosoky, G. and Mahmoud, M.E.F. [1993]: Lead pollution of water resources and its biochemical effects. Proc. 3rd. Int. Conf. on Environ. Prot. is a must, 75-87.
- [140] Adak. S.K. [1995]: Hematological studies on fresh water teleost, *Oreochromis* (*Tilapia*) *mosambicus* (Peters) in the normal conditions and under the stress of zinc and Arsenic. Ph.D. Thesis, zoology, Sambalpur Univ.
- [141] Shabana, M.B. [1963]: Induced pathological and Biological stress of acute lead poisoning in Egyptian cat fish *Clarieas Lazera*. Bull. Fac. Sci. Alex. Univ., 23: 1-141.
- [142] Ahmed, I.M. and S.Munshi [1992]: Scanning electron microscopic evaluation of changes on the morphology of blood cells of an Indian major carp *Clata catta* (Halm). Following exposure to copper. J. Environ. Biol. 13 [14]: 297-301.
- [143] Ghazaly, K.S. and K.M. Said (1995): Physiological characteristics of *Tilapia nilotica* under stress of copper. J. Egypt. Ger. Soc. Zool.
- [144] Arafa, M.M. and Ali, A.T. [2008]: Effect of some heavy metals pollution in Lake Mariout on *Oreochromis niloticus* fish Egypt. J. Comp. Path. & Clinic. Path., 21[3]: 191 – 201.
- [145] Yacoub, A.M. and Gad, N.S. [2012]: Accumulation of some heavy metals and biochemical alterations in muscles of *Oreochromis niloticus* from the River Nile in Upper Egypt. International Journal of Environmental and Science Education, 3:1-10.
- [146] Aboud, O. A.S.A. [2010]: Impact of pollution with lead, mercury and cadmium on the immune response of *Oreochromis niloticus*. New York Science Journal, 3[9]:12-16.
- [147] Gill, T.S. and Pant, J.C. [1985]: Erythrocytic and leukocytic responses to cadmium poisoning in a freshwater fish, *Puntius conchonus* Ham. Env. Res., 36 [2]: 327-337.
- [148]. Zaghoul, K.H.; Hasheesh, W.; Marie, M. and Zahran, I. [2007]: Ecological and biological studies on the Nile tilapia *Oreochromis niloticus* along different sites of lake Burullus. Egypt. J. Aquat. Biol. & Fish., 11[3]: 57 - 88.
- [149] Haggag, A.M.; Marie, M.A.S. and Zaghoul, K.H. [1999]: Seasonal effects of the industrial effluents on the Nile catfish; *Clarias gariepinus*. J. Egypt. Ger. Soc. Zool., 28[A]: 365-391.
- [150] Zaghoul, K.H. [2001]: Usage of zinc and calcium in inhibiting the toxic effect of copper on the Afriac catfish; *Clarias gariepinus*. J. Egypt Ger. Soc. Zool., 35[C]: 99- 120.
- [151] Mazon, A. F.; Monteiro, E. A. S.; Pinheiro, G. H. D. and Fernandes, M. N. [2002]: Gill cellular changes induced by copper exposure in the South American tropical freshwater fish *Prochilodus scrofa*. Environm. Res. A., 88: 52- 63.

- [152] Zaghoul, K.H.; Omar, W.A. and Abo-Hegab, S. [2005]: Environmental hazard risk assessment on; *Oreochromis niloticus* and *Tilapia zilli* fish. J. Egypt. Ger. Sco. Zool., 46[A]: 105- 139.
- [153]Ahmed, D.F.I. [2013]: Effect of industrial waste discharges including heavy metals in Borullus lake on some physiological parameters and antioxidants in *Tilapia niloticus* and *Siluriformes* fish. New York Science Journal,6 [4]:85-92.
- [154]Ahmad, I.; Oliveira, M.; Pacheco, M. and Santos, M.A. [2005]: *Anguilla anguilla* L. oxidative stress biomarkers responses to copper exposure with or without anaphthoflavone pre-exposure. Chemosphere, 61[2]: 267 – 275
- [155] Ercal, N.; Gurer-Orhan, H. and Aykin-Burns, N. [2001]: Toxic metals and oxidative stress part I: mechanisms involved in metal induced oxidative damage. Curr.Top. Med. Chem. 1: 529-539.
- [156]Doherty, V.F.; Ogunkuade, O. and Kanife, U.C. [2010]: Biomarkers of oxidative stress and heavy metal levels as indicators of environmental pollution in some selected fishes in Lagos, Nigeria. American-Eurasian J. Agric. & Environ. Sci., 7 [3]: 359-365.
- [157] Olagoke, O. [2008]: Lipid peroxidation and antioxidant defense enzymes in *Clarias gariepinus* as useful biomarkers for monitoring exposure to polycyclic aromatic hydro-carbons. MSc Theses, University of Lagos, Lagos, Nigeria.70.
- [158]Faramobi, E.O.; Adewole, A. and Ajimoko, Y.R. [2007]: Effect of butachlor on antioxidant enzyme status and lipid peroxidation in fresh water African Catfish, (*Clarias gariepinus*). Int. J. Environ. Res. Public Health, 5(5): 423-427.

الملخص العربي

المصادر والتأثيرات السمية للتلوث السطحي للمياه على الأسماك في مصر

أماني ثروت محمد خليل*، مجدي فكري ابو الفتوح، ميادة رجب فرج ، ياسمينا محمد عبد الحكيم، ولاء محمد عبد الهادي

قسم الطب الشرعي والسموم، كلية الطب البيطري، جامعة الزقازيق، مصر

تم تصنيف مصر كواحدة من أكثر البلدان التي يهددها نقص المياه بحلول عام ٢٠٢٥ بسبب الزيادة السكانية. أدى الترسيب الكبير للنفايات السائلة المنزلية والصناعية الملوثة في المياه إلى إلحاق ضرر مستمر وسريع بسطحها ومياهها الجوفية. في مصر ، المصدر الرئيسي للتلوث هو النفايات الصناعية ، حيث يتم تصريفها مباشرة في النيل أو من خلال النظام البلدي. من ناحية أخرى ، فإن تلوث البحر الأبيض المتوسط في الإسكندرية يرجع أساساً إلى المصانع الصناعية التي تصرف مياه الصرف الصحي في البحر عن طريق بحيرة ماريوت. كما ان سكان الأرياف لا يتمتعون بإمكانية الوصول إلى شبكات الصرف الصحي أو مرافق معالجة المياه العامة ، بل إنهم لا يحصلون عليها ، ولا يعتمدون إلا على التخلص من المياه المستعملة في نهر النيل في منطقة الدلتا ، يتم إعادة استخدام مياه الصرف للري بعد مزجها بمياه النيل و بعد إنشاء السد العالي ، تم استخدام الأسمدة والمبيدات الحشرية على نطاق واسع مما أدى إلى ازدهار الأعشاب الضارة التي تمنع الممرات المائية ، وتوفر عوائل لقواقع البلهارسيا. تتعرض الأسماك للسمية بسبب تلوث المياه من المصانع الكيماوية وأنبيب الصرف الصحي. وتتمثل المصادر الرئيسية لتلوث المياه المعادن الثقيلة و حرق الفحم والنفط وأسمدة الفوسفات واللدائن ومبيدات الآفات التي يتم القاءها في مياه الانهار والبحار و في الآونة الأخيرة ، لوحظت مخاطر محتملة لتسمم المعادن الثقيلة في الإنسان بسبب استهلاك الأسماك الملوثة